Review articles

Anisakis spp. as etiological agent of zoonotic disease and allergy in European region – an overview

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ABSTRACT. Nematodes of the genus *Anisakis* are worldwide distributed marine species parasitized many fish and cephalopod species as larvae and sea mammals as adult form. Anisakiosis as food-borne disease is an important public health problem worldwide. Human become infected by eating raw or undercooked fish or squids. Well documented are gastrointestinal response to infection but increasingly allergic symptoms were observed also after eating well cooked fish. This is because some of allergens of *Anisakis* are thermostable and resistant to pepsin treatment. Due to a significant increase in human mobility and global transport of fresh products like fish on ice, food-borne diseases require educational campaigns that pay attention to threats in various parts of the world.

Keywords: anisakiosis, food-borne diseases, allergy, emergent zoonosis, health education

Introduction

Parasites are natural and significant components of all ecosystems. Many parasites have a complex life cycle and entering into interactions with invertebrate and vertebrate hosts [1], also humans. The increasing importance of zoonoses, especially food-borne diseases, transmitted between vertebrate animals and humans, in the public health have been observed. The popularity of raw fish in the recent years is growing considerably in many countries, becoming a new culinary habit. Raw fish poses some potential hazards for consumers, also parasitic diseases and allergy. In good American and European sushi bar dishes has been prepared properly according to regulations by the U.S. Food and Drug Administration (FDA) and the European Food Safety Authority (EFSA) which minimizes the risk of infection. Sometimes people prepare raw or lightly cooked fish dishes at home and they don't known that they must use frozen fish, not fresh. Globalization causes that people more travelling, and also food from all over the world is available in markets but people don't know the regional threats.

Anisakiosis is one of the food-borne parasitic diseases transmitted by fish and some cephalopods.

The agents are nematodes from the family Anisakidae, most commonly from genera Anisakis, Pseudoterranova and Contracaecum [2,3]. Anisakis can cause not only parasitic disease but also allergy. The ingestion infected fish products with a live larva of Anisakis can cause a various symptoms such as gastric, intestinal or allergic reaction and with death larva various allergic symptoms. Currently the number of both, infection and allergy, have increased worldwide [4], but the exact number of gastrointestinal anisakiosis cases and allergic reaction is unknown due to lack of epidemiological data. EFSA reported about 20 000 cases of anisakiosis worldwide before 2010, where of 90% concerns Japan. In Europe, most cases are reported from Spain, Italy and less from the Netherlands and Germany [5–7].

The prevalence of allergic disease, in this food allergy, increased worldwide and become an important public health problem [8]. Some parasitic infection or contact with parasite proteins associated with allergic reaction *e.g.* food allergy, asthma like syndrome, urticaria, anaphylaxis. Some people think that have allergies to fish flesh and thus they are forced to eliminate all fish from their diet. But sometimes the allergic reaction may not be to the fish protein but to the fish parasites. *Anisakis* produces 14 substances that may provoke allergies in humans and several novel probable allergens [9,10]. Some of antigens are thermostable therefore allergic reaction occurred also after consumption of heat treated fish. If allergy testing confirm only reaction to *Anisakis* antigens and rule out to fish, patient must avoid in his diet only fish species known as *Anisakis* hosts and *i.e.* just to be sure, eat only freshwater fish. Because in markets are available fresh fish from the various fishing grounds around the world it is important to share information about hosts, appropriate cooking methods and potential risk of allergy caused by *Anisakis*.

The aim of this paper is to draw attention of the public health specialists to need resource for about the problem of anisakiosis worldwide as well as food allergy, and the most effective ways to risk avoidance.

Biology – distribution and life cycle of *Anisakis*

Nematodes of the genus *Anisakis* are marine species with worldwide distribution. Nowadays, based on the molecular methods, genus *Anisakis* includes nine species: *A. simplex sensu stricto*, *A. pegreffii*, *A. berlandi* (=*A. simplex* sp. C), *A. typica*, *A. nascettii*, *A. paggiae*, *A. physeteris*, *A. brevispi-culata* and *A. ziphidarum* [11–16]. These species are morphologically similar but genetically different. They have also different distribution, life cycle and host preferences [11–13].

The most important for public health are *A.* simplex s.s. distributed in northern hemisphere, especially North Atlantic, *A. pegreffii* in central Atlantic and Mediterranean Sea, *A. physeteris* in Atlantic and Mediterranean and *A. berlandi* in Pacific and Antarctic regions [12,13,17].

According to Klimpel et al. [14,18,19] the life cycle of *Anisakis* is sophisticated and still poorly understood because of low host specificity and modifications dependent on the geographical region and feeding ecology of local animal community.

Anisakis is a nematode parasitizes alimentary tract, particularly stomach, of cetaceans like dolphins, porpoises and whales [12,13,20–23]. A total of 53 sea mammals species worldwide were known as final hosts [18]. In the European waters adult nematodes have been noted mainly from harbour porpoise (*Phocoena phocoena*) [21,22, 24–27], common bottlenose dolphin (*Tursiops*)

truncatus) [18,23,24], white-beaked dolphin (*Lagenorhynchus albirostris*) [24], common minke whale (*Balaenoptera acutorostrata*) and long-finned pilot whale (*Globicephala melas*) [18,23]. Harbour porpoise act as local final host in the North Sea and the Baltic Sea [18,25]. After Smith and Wootten [24], sometimes, but less frequently *Anisa kis* were noted in pinnipeds. Skrzypczak et al. [28] found small number of L3 and L4 of *A. simplex*, but no adults, in grey seal (*Halichoerus grypus*) and common seal (*Phoca vitulina*) from the southern Baltic Sea and they suggests that seals are rather accidental hosts.

Eggs produced by adult females in the alimentary tract of the final host are shed into the water where embryonated and develop. Nagasawa [29] described free-living L2, but now, it is known that one or two molts occur within the egg and L2 or L3 hatch to the environment and free-living [30]. Marine crustaceans feeding on free-living stages, L2 or L3, became infected. After ingestion larvae migrate to the haemocoel. Fish are infected with eating infected crustaceans or other smaller fish. In fish organism larvae penetrate intestinal wall and located on the pyloric caeca, mesentery of intestine, liver or gonads. Some of larvae migrate to the muscles [15]. Anisakis has been noted from more than 200 fish and 25 cephalopods throughout the world [18]. When the infected fish is eaten by a sea mammal, the life cycle is closed.

Human [31], some fish eating mammals like brown bear (*Ursus arctos*) [20], river otter (*Lutra canadensis*) [32], Eurasian otter (*Lutra lutra*) [33], as well as birds like great cormorants (*Phalacrocorax carbo*) [34,35], northern fulmar (*Fulmarus glacialis*) [2,26,37], Atlantic puffin (*Fratercula arctica*), Brünnich's guillemot (*Uria lomvia*) [37] or common gull (*Larus canus*), [cf. 34] are known as accidental hosts.

European fish-hosts and public health

The third stage larvae of *Anisakis* infected the viscera and/or the musculature of many fish species and some cephalopods [13,14]. *Intra vitam* larval migration can occur [38,39] and sometimes also *post mortem* migration is possible [38,40,41]. Because of worldwide distribution of these nematodes and global transport of fresh fish on the ice to the markets in distant regions makes anisakiosis an emerging public health problem.

In both, Atlantic and Pacific regions, wild

salmons species are commonly infected with nematodes [15,42,43]. According to some papers farmed salmons are free from anisakid larvae because of fact that they are feed on processed food [*e.g.* 42,44,45]. Nonetheless a very low risk of infection after consuming of raw farmed salmons also exists. Farmed marine fish such as salmons are cultured in open cages and can be infected by parasites *via* intermediate host entering the cages or free stages in water. Mo et al. [46] found *A. simplex* larvae in farmed Atlantic salmons in Norway. Certainly nematodes in farmed fish occurred less frequently than in wild salmonids.

In European waters fish like the European hake (Merluccius merluccius) [47,48], European anchovy (Engraulis encrasicholus) [3,41,49,50], European pilchard (Sardina pilchardus) [50], Atlantic mackerel (Scomber scombrus) [48,51], horse mackerel (Trachurus trachurus) [48,52], blue whiting (Micromesistius poutassou) [48,53] are a common hosts. Silva and Eiras [48] also found larvae of Anisakis in European pilchard (Sardina pilchardus), pouting (Trisopterus luscus) and less in black seabream (Spondyiliosoma cantharus), sole (Solea sp.) and tub gurnard (Chelidonichthys lucerna). In the Atlantic and the North Sea some commercial fish species, like herring (Clupea harengus), saithe (Pollachius virens), cod (Gadus morhua), redfish (Sebastes marinus) [18,54], and also European sprat (Sprattus sprattus) [55] are infected with larvae of A. simplex. Also larvae were noted in small fish pearlside (Maurolicus muelleri) without commercial importance but ecologically valid [18].

In the Baltic Sea the most infected fish species is herring (*Clupea harengus membras*) [*e.g.* 56–59]. Less frequently *Anisakis* infected cod [59–64]. Sporadically nematodes occurred in other fish species like flounder (*Platichthys flesus*) [59,62], zander (*Sander lucioperca*) [65] or garfish (*Belone belone*) [66].

Occasionally *Anisakis* infected gobies [67,68] and three-spined stickleback (*Gasterosteus acule-atus*) [69]. Experiments conducted by Køie [70] as well Rolbiecki et al. [71] showed that larvae from herring viscera are infective if were ingested by other fish, like three-spined stickleback or flounder. European sprat noted as host in the Atlantic and the North Sea, in the Baltic Sea is not infected [55,59].

Prevalence of *A. simplex* in the Baltic herring, and also in cod decreased from south-western to the south-eastern Baltic fishing grounds [59,64]. In the

Baltic Sea many populations of herrings are living [72] and seasonal fluctuation in the occurrence of A. simplex was observed. Infected specimens of herring appeared from autumn to spring [59,73]. The Baltic herrings from spring coastal stock were infected with intensity from a few to hundreds larvae Anisakis per fish located mostly on mesentery, pyloric caeca, gonads but also in the muscles [59]. Herring infection increased with fish age and size. Herring 1-2 years old are free of nematodes and oldest fish accumulated parasites [73-75]. Podolska [75] found parasites only in herring more than 21 cm in length and herrings up to 20 cm in length are free from parasites. Really similar correlation occurred also in case of Baltic cod [59,64] as well as sardines in Spain [76].

Anisakiosis – human infection

Human is not a natural host, but can be an accidental host for Anisakis. The larvae are able to survive in humans a few days up to weeks [49]. Humans acquire infection by eating raw fish dishes, e.g. sushi, sashimi, ceviche, anchovies marinated in vinegar anchovies, gravlax, lomi-lomi, herring roe and also inadequately cooked fish or squids [31]. This fish-borne disease was first described in the 1960s in the Netherlands [49,77,78]. It was a result of consumption of "green" herring, very popular in this country, lightly salted herrings. Nowadays cases of infection are reported worldwide [49,79], in Asia countries as Japan, Korea, Taiwan, China, Malaysia [10,49,80-83], in Europe, as Spain, Italy, the Netherlands, Denmark, Germany, France, United Kingdom, Norway and Croatia, in North America noted in Canada and the United States of America with Alaska and Hawaii, in South America countries like Brazil or Chile [9,10,49,80-83], in Africa in Egypt [49], South Africa [84], Morocco [85,86] also in Australia [87] and New Zealand [49].

In the European Union totally 236 cases of anisakiosis were noted from 2000 to 2016. Most of them were detected in Spain 66.9% and Italy 28.4%. In 62 cases, infection was observed after ingesting European anchovy (*Engraulis encrasicholus*). The main symptoms that have been noted include itching, abdominal pain and cutaneous symptoms, less frequently vomiting, diarrhea, fever and pain in the chest. In most cases, the diagnosis was based on serological tests (86.4%) and gastroscopy (10.2%) [88]. Nowadays Spain is second, after Japan country with highest number of cases of anisakiosis [89].

Well documented are gastrointestinal response to infection but nowadays more often allergic reaction to Anisakis proteins were observed also after eating properly cooked fish infected with A. simplex [9,49]. Nematodes can parasitize humans but never mature. Larvae penetrate gastrointestinal mucosa or rarely other organs with clinical manifestation, also can cause allergic reaction, i.a. urticaria and angioedema [31,90]. Despite this anisakiosis is a relevant zoonotic disease especially since the increased popularity of sushi bar in the world. A human health risk is after eating traditional dishes from raw or low salted or marinated fish like sushi, sashimi, green herring, ceviche, gravlax, salmon tartare, lomi-lomi, anchovies marinated in vinegar or other regional cuisine [5,31,77,91,92]. In the United States, anisakiosis reported mostly after eating of pacific salmons (e.g. Oncorhynchus nerka, O. keta) [15,49,92]. In the Western Europe it is herring. Eating of flounder or sprat practically does not health hazard [59]. At the beginning of the 21st century prevalence of Anisakis in cod was low and risk for health was very low but nowadays infection clearly increasing [64,93]. In Spain, Portugal, France, Italy and Croatia it is pickled anchovies (Engraulis encrasicholus) [49] and raw sardines European pilchards (Sardina pilchardus) [10]. Also European hake (Merluccius merluccius) [47], Atlantic mackerel (Scomber scombrus) [51] or horse mackerel (Trachurus trachurus) [52], salmon (Salmo salar) [10] represents a risk of Anisakis infection.

Eating raw, lightly salted or cold smoked huge specimens of Baltic herring or their roe carries a health risk from autumn to spring. Every year, during this time larvae of *Anisakis* in herrings, mostly 25 cm in length and above, both fresh and smoked, in the fish markets were observed. These nematodes were found in smoked herring by Guz et al. [94]. Szostakowska et al. [59] found larvae, also alive, in preserves from herring – smoked and marinated. Bilska-Zając et al. [95] documented *Anisakis* larvae in frozen salmon filets from markets. Spirally convoluted third stage larvae of *A. simplex*, lived in the viscera, gonads or muscles of fish and cephalopod are visible to the naked eye.

Clinical manifestation

Gastrointestinal infection

Parasites located in the gastric or intestinal mucosa. Consumption raw or undercooked fish with

living larvae provoke local irritation, severe epigastric pain, nausea, vomiting, diarrhoea, lowgrade fever. Symptoms usually appear up to 12 hour after fish consumption in the case of gastric infection and 5-7 days in the case of intestinal infection. Mucosal oedema around the area of penetration is observed and also eosinophilic intestinal lesion or perforation sometimes occurs. Symptoms may be similar to i. a. gastric or duodenal ulcer, appendicitis or peritonitis. Because symptoms are not specific patients with anisakiosis sometimes were misdiagnosed [10,49,96-98]. According to Guardone et al. [90] pathologists should consider the possibility of anisakiosis when eosinophilic granuloma of the gastrointestinal tract, the mesentery or the peritoneum occurred. The gastric or intestinal infections may be accompanied by allergic reaction like urticaria, angioedema or also anaphylactic shock [10,49,96.97].

The diagnosis is based on endoscopy, laparotomy or radiological examination and various immunological tests [113]. Endoscopic extraction of *Anisakis* is the most common method of treatment. Small bowel infection may be treated also with albendazole therapy [49,99,100].

Allergy

Kasuya et al. [101] described for the first time *A.* simplex allergy in Japan. Since then, many cases of allergy were noted worldwide, especially in Spain [*i.e.* 10,49,102,103]. Allergy may be accompanied by invasion but also was observed after eating properly cooked fish and canned fish – exposure to allergenic proteins in food [5,10,104]. Clinical symptoms were also observed in patients after eating frozen fish [5,105,106]. According to Moneo et al. [107] in endemic countries more than 7% of population could be sensitized.

Anisakis can cause to induce IgE-mediated reactions with clinical symptoms as urticaria, angioedema, asthma and anaphylaxis after consumption of parasitized fish [4,108]. Some cases of allergy were diagnosed also after eating meat of chicken feeding on fishmeal, which contained nematodes [109]. Cases of occupational allergy were also noted after contact or inhalation of *Anisakis* allergens. Hypersensitivity reactions as contact dermatitis, conjunctivitis and asthma in fishermen, fishmongers and a chicken breeder were demonstrated after contact with infected fish or fishmeal [84,109–112].

The allergic reaction described for anisakiosis

includes urticaria, angioedema, in this facial angioedema, gastrointestinal reaction as nausea, vomiting and/or diarrhea and also respiratory reaction. Anaphylactic shock and respiratory arrest were also noted [*i.e.* 10,49,102,103].

Anisakis produces 14 proteins that may provoke allergies in humans accepted in the International Union of Immunological Societes and published in website allergen.org [10]. Several novel probable allergens also were noted [9,10]. In allergome database were noted 46 items [www.allergome.org]. Allergic proteins of Anisakis are classified to tree groups:

- Excretory/secretory proteins expressed by larvae during infestation as Ani s 1, Ani s 4–9,
- Somatic proteins constituents of the larvae body as Ani s 2, Ani s 3,
- Cuticular proteins occurred on the body surface of larvae and protect them from digestion [9,98,113].

Some of allergens of *Anisakis* are thermostable, show resistance to pepsin digestion and low pH therefore allergic reaction is possible also after eating dead larvae with fish flesh for example after conventional or microwave heat treatment [114–116]. Allergic reaction occurred also after freezing fish products [115]. Humans are primarily exposed to somatic and cuticular antigens also from death larvae contained in food and excretory/ secretory antigens when alive larvae penetrate tissue. Allergy recognized by more than 50% patients was most often caused by allergens Ani s 1, Ani s 2 and Ani s 7 [98,113,117,118].

In diagnosis are used the skin prick test and the detection of specific IgE using antigen from nematode, but they are interfered with crossreactivities of other ascarides, mites, shrimps and some microorganism [107,114,119]. Ivanovic et al. [98] also lists method like complement fixation test (CFT), immunofluorescent-antibody test (IFAT), immunodiffusion test (IDT), immunoelectrophoretic assays, enzyme-linked immunosorbent assay (ELISA). According to Lorenzo et al. [120] and Carballeda-Sangiao et al. [121] the fluorescence enzyme immunoassay (CAP FEIA) is the most used method for laboratory diagnosis of Anisakis allergy because of high sensitivity and low specificity. The major problem in diagnosis of allergic reactions is the presence of cross-reactivity of Anisakis allergens e.g. tropomyosin, paramyosin, with antigens of other nematodes, insects, mites, crustacean and mollusks [10].

Prevention

A. simplex infection may pose a health risk if people eat raw, undercooked, pickled and coldsmoked fish dishes. Parasite infection is not a health hazard when the fish or other seafood is thoroughly cooked but allergy is possible because of thermostable allergens. How Moneo et al. [107] concluded, information about potential health risk of consumption habits would avoid contacts with parasite allergens and consequently decrease the appearance of acute or chronic episodes induced by the parasites.

The best preventive measure on anisakiosis is educating people about the risk of eating raw, inadequately cooked, lightly pickled or salted marine fish or squids, also information about the fish species carrying the greatest risk of disease.

The risk of human infection can be reduced:

- the fresh fish as soon as possible should be carefully eviscerated and wash;
- visual examination of fish: extraction of visible parasites and elimination of heavily parasitized fish – this method is also applicable at home;
- visual inspection of the whole fish abdominal cavity used as standard method in the European Union does not guarantee larvae detection, especially when they can migrate to the fish muscles [117,122]. Presence of Anisakidae larvae in fish products can be assessed by various, more effective laboratory methods, i.a. direct observation of body cavity, liver and gonads, compressor method with illuminated table and UV transillumination detected parasites in muscles, or chlorine-peptic digestation of samples of muscle tissue, similar to methods of detecting *Trichinella* in muscles of pigs, wild boars and other animals [122–124].
- thermal processing heating or freezing. The European Food Safety Authority (EFSA) and the USA Food and Drug Administration (FDA) recommends:
 - cooking fish at > 60°C (> 145°F) for 15 seconds is sufficient to kill parasites. This means that the common fish fillet must be heat treated for about 10 minutes;
 - fish for raw or lightly cooked consumption must be freezing at -20°C (-4°F) for not less than 24 h or at least four days in a domestic freezer at -15°C (EFSA). FDA recommended freezing at -20°C (-4°F) for at least 7 days or blast freezing to - 35°C (-31°F) for ≥15 h [6,125]. This recommendation prevent

gastrointestinal infection, but the study of Rodriguez-Mahillo et al. [115] indicates that the antigenicity of *Anisakis* larvae is preserved after freezing and may sensitized humans after ingestion of infected frozen fish.

More restrictive FDA recommendation is a result of the presence of more resistant *Pseudoterranova* in the North America. European recommendation should also be tightened because the prevalence of fish, like cod, infected with Pseudoterranova increased in the last twenty years, also in the Baltic Sea [64,93,126]. Pseudoterranova like Anisakis can cause both, gastrointestinal infection and allergic reaction in humans [127,128]. However, the allergic potencial of other Anisakidae nematodes, such as Pseudoterranova and Contracacecum, has not yet been well studied and requires further research [129]. In addition, the latest research carried out by Podolska et al. [130] on freezing cod fillets and whole herrings and the survival rate of nematode larvae have shown that both, Pseudoterranova and Anisakis, died in cod fillets at a temperature -15°C or lower but in whole herring some Anisakis larvae survived at -20°C. These results demonstrate that in the freezing procedure the capability of the freezing device, as well the thickness and type of the fish products being frozen are important [130].

In conclusion, prevention of fish-borne disease requires monitoring of parasites, early fish evisceration and control fish products [122-124]. Very important are education campaigns about home preparation methods, especially fish freezing capacity and properly cooking techniques, also widely available information about hosts of parasites. It is important to known that visual inspection of the whole fish abdominal cavity, possible to use at home, does not guarantee larvae detection. In addition properly cooking or freezing fish products can protect from gastrointestinal infection but not allergy because some proteins have been heat, freezing and pepsin resistant e.g. Ani s 1, Ani s 4, Ani s 5, Ani s 8, Ani s 9, Ani s 10, Ani s 11.0201 [104,114-116,129]. Allergy cases after consuming marine fish (raw, stewed, fried, frozen, smoked, marinated, canned) should be verified by an allergist, whether it is an allergy to fish protein or parasites.

References

[1] Marcogliese D.J. 2005. Parasites of the super-

organism: Are they indicators of ecosystem health? *International Journal for Parasitology* 35: 705-716. doi:10.1016/j.ijpara.2005.01.015

- [2] Lymbery A.J., Cheah F.Y. 2007. Anisakid nematodes and anisakiosis. In: *Food Borne Parasitic Zoonosis: Fish and Plant-Borne Parasites* (Eds. K.D. Murrell, B. Fried). Springer, New York, USA: 185-207. doi:10.1007/978-0-387-71358-8_5
- [3] De Liberato C., Bossů T., Scaramozzino P., Nicolini G., Ceddia P., Mallozzi S., Cavallero S., D'Amelio S. 2013. Presence of anisakid larvae in the European Anchovy, *Engraulis encrasicolus*, fished off the Tyrrhenian Coast of Central Italy. *Journal of Food Protection* 76: 1643-1648.

doi:10.4315/0362-028X.JFP-13-092

- [4] Lin A.H., Nepstad I., Florvaag E., Egaas E., Van Do T. 2014. An extended study of seroprevalence of anti-*Anisakis simplex* IgE antibodies in Norwegian blood donors. *Scandinavian Journal of Immunology* 79: 61-67. doi.org/10.1111/sji.12130
- [5] Audicana M.T., Ansotegui I.J., de Corres L.F., Kennedy M.W. 2002. Anisakis simplex: dangerous dead and alive? Trends in Parasitology 18: 20-25. doi:10.1016/S1471-4922(01)02152-3
- [6] EFSA Panel on Biological Hazards (BIOHAZ); Scientific Opinion on risk assessment of parasites in fishery products. EFSA Journal 2010; 8(4):1543. [91 pp.]. doi:10.2903/j.efsa.2010.1543
- [7] Bao M., Pierce G. J., Pascual S., González-Muňoz M., Mattiucci S., Mladineo I., Cipriani P., Bušelić I., Strachan N.J.C. 2017. Assessing the risk of an emerging zoonosis of worldwide concern: anisakiasis. *Scientific Reports* 7, [43699]. https://doi.org/10.1038/srep43699
- [8] Del Rey Moreno A., Valero A., Lozano Maldonado J., Mayorga C., Gómez B., Torres M.J., Hernández J., Ortiz M., Lozano Maldonado J. 2006. Sensitization to *Anisakis simplex* s.l. in a healthy population. *Acta Tropica* 97: 265-269. doi:10.1016/j.actatropica.2005.11.007
- [9] Fæste C.K., Jonscher K.R., Dooper M.M.W.B., Egge-Jacobsen W., Moen A., Daschner A., Egaas E., Christians U. 2014. Characterisation of potential novel allergens in the fish parasite *Anisakis simplex*. *EuPA Open Proteomics* 4:140-155. doi:10.1016/j.euprot.2014.06.006
- [10] Aibinu I., Smooker P.M., Lopata A.L. 2019. Anisakis nematodes in fish and shellfish – from infection to allergies. *IJP: Parasites and Wildlife* 9: 384-393. doi:10.1016/j.ijppaw.2019.04.007
- [11] Valentini A., Mattiucci S., Bondanelli P., Webb S.C., Mignucci-Giannone A.A., Colom-Llavina M.M., Nascetti G. 2006.Genetic relationships among *Anisakis* species (Nematoda: Anisakidae) inferred from mitochondrial cox2 sequences, and comparison with allozyme data. *Journal of Parasitology* 92: 156-166. doi:10.1645/GE-3504.1

[12] Mattiucci S., Nascetti G. 2006. Molecular systematics, phylogeny and ecology of anisakid nematodes of the genus *Anisakis* Dujardin, 1845: an update. *Parasite* 13: 99-113. doi:10.1051/acmeits/20006122000

doi:10.1051/parasite/2006132099

[13] Mattiucci S., Nascetti G. 2008. Advances and trends in the molecular systematics of anisakid nematodes, with implications for their evolutionary ecology and host-parasite co-evolutionary processes. *Advances in Parasitology* 66: 47-148.

doi:10.1016/S0065-308X(08)00202-9

[14] Klimpel S., Kellermanns E., Palm H.W. 2008. The role of pelagic swarm fish (Myctophidae: Teleostei) in the oceanic life cycle of *Anisakis* sibling species at the Mid-Atlantic Ridge, Central Atlantic. *Parasitology Research* 104: 43-53.

doi:10.1007/s00436-008-1157-3

- [15] Karl H., Baumann F., Ostermeyer U., Kuhn T., Klimpel S. 2011. Anisakis simplex (s.s.) larvae in wild Alaska salmon: no indication of post-mortem migration from viscera into flesh. Diseases of Aquatic Organisms 94: 201-209. doi:10.3354/dao02317
- [16] Mattiucci S., Cipriani P., Webb S.C., Paoletti M., Marcer F., Bellisario, B., Gibson D.I., Nascetti G. 2014. Genetic and morphological approaches distinguish the three sibling species of the *Anisakis simplex* species complex, with a species designation as *Anisakis berlandi* n. sp. for *A. simplex* sp. C (Nematoda: Anisakidae). *Journal of Parasitology* 100: 199-214. doi:10.1645/12-120.1
- [17] Klimpel S., Palm H.W. 2011. Anisakid nematode (Ascaridoidea) life cycles and distribution: Increasing zoonotic potential in the time of climate change? In: *Progress in Parasitology* (Ed. H. Mehlhorn). *Parasitology Research Monographs* vol. 2. Springer Verlag, Berlin, Heidelberg: 201-222. doi:10.1007/978-3-642-21396-0 11.
- [18] Klimpel S., Palm H.W., Rückert S., Piatkowski U. 2004. The life cycle of *Anisakis simplex* in the Norwegian Deep (northern North Sea). *Parasitology Research* 94: 1-9. doi:10.1007/s00436-004-1154-0
- [19] Klimpel S., Kuhn T., Busch M.W., Karl H., Palm H.W. 2011. Deep water life cycle of *Anisakis paggiae* (Nematoda: Anisakidae) in the Irminger Sea indicates kogiid whale distribution in north Atlantic waters. *Polar Biology* 34: 899-906. doi:10.1007/s00300-010-0946-1
- [20] Davey J.T. 1971. A revision of the genus Anisakis Dujardin, 1845 (Nematoda: Ascaridata). Journal of Helminthology 45: 51-72. doi:10.1017/S0022149X00006921
- [21] Herreras M.V., Kaarstad S.E., Balbuena J.A., Kinze C.Chr., Raga J.A. 1997. Helminth parasites of the digestive tract of the harbour porpoise *Phocoena phocoena* in Danish waters: a comparative geographical analysis. *Diseases of Aquatic Organisms* 28: 163-167.

- [22] Herreras M.V., Balbuena J.A., Aznar F.J., Kaarstad S.E., Fernández M., Raga J.A. 2004. Population structure of *Anisakis simplex* (Nematoda) in harbor porpoises *Phocoena phocoena* off Denmark. *Journal* of *Parasitology* 90: 933-938. doi:10.1645/GE-188R
- [23] Ugland K.I., Strømnes E., Berland B., Aspholm P.E. 2004. Growth, fecundity and sex ratio of adult whaleworm (*Anisakis simplex*; Nematoda, Ascaridoidea, Anisakidae) in three whale species from the North-East Atlantic. *Parasitology Research* 92: 484-489. doi:10.1007/s00436-003-1065-5
- [24] Smith J.W., Wootten R. 1978. Anisakis and anisakiosis. Advances in Parasitology 16: 93-163. doi:10.1016/S0065-308X(08)60573-4
- [25] Siebert U., Wünschmann A., Weiss R., Frank H., Benke H., Frese K. 2001. Post mortem findings in harbour porpoises (*Phocoena phocoena*) from the German North and Baltic Seas. *Journal of Comparative Pathology* 124: 102-114. doi:10.1053/jcpa.2000.0436
- [26] Wunschmann A., Siebert U., Frese K., Weiss R., Lockyer C., Heide-Jřrgensen M. P., Müller G., Baumgärtner W. 2001. Evidence of infectious diseases in harbour porpoises (*Phocoena phocoena*) hunted in the waters of Greenland and by-caught in the German North Sea and Baltic Sea. Veterinary Record 148: 715-720. doi:10.1136/vr.148.23.715
- [27] Lehnert K., Seibel H., Hasselmeier I., Wohlsein P., Iversen M., Nielsen N.H., Heide-Jørgensen M.P., Prenger-Berninghoff E., Siebert U. 2014. Increase in parasite burden and associated pathology in harbour porpoises (*Phocoena phocoena*) in West Greenland. *Polar Biology* 37: 321-331. doi:10.1007/s00300-013-1433-2
- [28] Skrzypczak M., Rokicki J., Pawliczka I., Najda K., Dzido J. 2014. Anisakids of seals found on the southern coast of Baltic Sea. *Acta Parasitologica* 59: 165-172. doi:10.2478/s11686-014-0226-2
- [29] Nagasawa K. 1990. The life cycle of Anisakis simplex. A review. In: Intestinal Anisakiosis in Japan. Infected Fish, Sero-Immunological Diagnosis, and Prevention (Eds. H. Ishikura, K. Kikuchi). Springer Verlag Tokyo: 31-40. doi:10.1007/978-4-431-68299-8 4
- [30] Køie M., Berland B., Burt M.D.B. 1995. Development to third stage larvae occurs in the eggs of Anisakis simplex and Pseudoterranova decipiens (Nematoda, Ascaridoidea, Anisakidae). Canadian Journal of Fisheries and Aquatic Sciences 52 (Suppl. 1): 134-139. doi:10.1139/f95-519
- [31] Chai J.Y., Darwin Murrell K., Lymbery A.J. 2005. Fish-borne parasitic zoonoses: status and issues. *International Journal for Parasitology* 35: 1233-1254. doi:10.1016/j.ijpara.2005.07.013
- [32] Hoberg E.P., Henny C.J., Hedstrom O.R., Grove R.A. 1997. Intestinal helminths of river otters (*Lutra canadensis*) from the Pacific northwest. *Journal of*

- [33] Torres J., Feliu C., Fernández-Morán J., Ruíz-Olmo J., Rosoux R., Santos-Reis M., Miquel J., Fons R. 2004. Helminth parasites of the Eurasian otter *Lutra lutra* in southwest Europe. *Journal of Helminthology* 78: 353-359. doi:10.1079/JOH2004253
- [34] Kanarek G., Rolbiecki L. 2006. Third-stage larvae of Anisakis simplex (Rudolphi, 1809) in the Great Cormorant [Phalacrocorax carbo sinensins (Blumenbach, 1798)] from the Vistula Lagoon, Poland. Oceanological and Hydrobiological Studies 35: 23-28.
- [35] Kanarek G., Zaleśny G. 2014. Extrinsic- and intrinsic-dependent variation in component communities and patterns of aggregations in helminth parasites of great cormorant (*Phalacrocorax carbo*) from N.E. Poland. *Parasitology Research* 113: 837-850. doi:10.1007/s00436-013-3714-7
- [36] Riley J. 1972. The pathology of *Anisakis* nematode infections of the Fulmar *Fulmarus glacialis*. *Ibis* 114: 102-104. doi:10.1111/j.1474-919X.1972.tb02594.x
- [37] Ólafsdóttir D., Lilliendahl K., Sólmundsson J. 1996. Nematode infections in icelandic seabirds. *Bulletin of the Scandinavian Society for Parasitology* 6: 124-125.
- [38] Smith J.W. 1984. The abundance of Anisakis simplex L3 in the body-cavity and flesh of marine teleosts. International Journal for Parasitology 14: 491-495. doi:10.1016/0020-7519(84)90030-4
- [39] Karl H. 2008. Nematode larvae in fish on the German market: 20 years of consumer related research. Archiv für Lebensmittelhygiene 59: 107-116. doi:10.2376/0003-925X-59-107
- [40] Smith J.W., Wootten R. 1975. Experimental studies on the migration of *Anisakis* sp. larvae (Nematoda: Ascaridida) into the flesh of herring, *Clupea harengus* L. *International Journal for Parasitology* 5: 133-136. doi:10.1016/0020-7519(75)90019-3
- [41] Cipriani P., Acerra V., Bellisario B., Sbaraglia G.L., Cheleschi R., Nascetti G., Mattiucci S. 2016. Larval migration of the zoonotic parasite *Anisakis pegreffii* (Nematoda: Anisakidae) in European anchovy, *Engraulis encrasicolus*: Implications to seafood safety. *Food Control* 59: 148-157. doi:10.1016/j.foodcont.2015.04.043
- [42] Food and Agriculture Organization of the United Nations (FAO). 2016. The state of world fisheries and aquaculture, 2016: contributing to food security and nutrition for all. FAO, Rome. http://www.fao.org/3/a-i5555e.pdf
- [43] Senos M., Poppe T., Hansen H., Mo T.A. 2013. Tissue distribution of Anisakis simplex larvae (Nematoda; Anisakidae) in wild Atlantic salmon, Salmo salar, from the Drammenselva river, south-east Norway. Bulletin of the European Association of Fish Pathologists 33: 111-117.
- [44] Angot V., Brasseur P. 1993. European farmed

Atlantic salmon (*Salmo salar* L.) are safe from anisakid larvae. *Aquaculture* 118: 339-344. doi:10.1016/0044-8486(93)90468-E

- [45] Lunestad B.T. 2003. Absence of nematodes in farmed Atlantic salmon (*Salmo salar* L.) in Norway. *Journal of Food Protection* 66: 122-124. doi:10.4315/0362-028X-66.1.122
- [46] Mo T.A., Gah A., Hansen H., Hoel E., Øaland Ø., Poppe T.T. 2014. Presence of Anisakis simplex (Rudolphi, 1809 det. Krabbe, 1878) and Hysterothylacium aduncum (Rudolphi, 1802) (Nematoda; Anisakidae) in runts of farmed Atlantic salmon, Salmo salar L. Journal of Fish Diseases 37: 135-140. doi:10.1111/jfd.12096
- [47] Daschner A., Alonso-Gómez A., Caballero T., Barranco P., Suarez-De-Parga J.M., López-Serrano M.C. 1998. Gastric anisakiosis: an underestimated cause of acute urticaria and angio-oedema? *British Journal of Dermatology*139: 822-828. doi:10.1046/j.1365-2133.1998.02507.x
- [48] Silva M.E.R., Eiras J.C. 2003. Occurrence of *Anisakis* sp. in fishes off the Portuguese West coast and evaluation of its zoonotic potential. *Bulletin of the European Association of Fish Pathologists* 23: 13-17.
- [49] Audicana M.T., Kennedy M.W. 2008. Anisakis simplex: from obscure infectious worm to inducer of immune hypersensitivity. Clinical Microbiology Reviews 21: 360-379. doi:10.1128/CMR.00012-07
- [50] Serracca L., Battistini R., Rossini I., Carducci A., Verani M., Prearo M., Tomei L., De Montis G., Ercolini C. 2014. Food safety considerations in relation to *Anisakis pegreffii* in anchovies (*Engraulis encrasicolus*) and sardines (*Sardina pilchardus*) fished off the Ligurian Coast (Cinque Terre National Park, NW Mediterranean). *International Journal of Food Microbiology* 190: 79-83. doi:10.1016/j.ijfoodmicro.2014.08.025
- [51] Pekmezci G.Z. 2014. Occurrence of Anisakis simplex sensu stricto in imported Atlantic mackerel (Scomber scombrus) represents a risk for Turkish consumers. International Journal of Food Microbiology 185: 64-68.
- doi:10.1016/j.ijfoodmicro.2014.05.018 [52] Adroher F.J., Valero A., Ruiz-Valero J., Iglesias L.
- [52] Adroner P.J., Valeto A., Ruiz-Valeto J., Igresias E. 1996. Larval anisakids (Nematoda: Ascaridoidea) in horse mackerel (*Trachurus trachurus*) from the fish market in Granada (Spain). *Parasitology Research* 82: 253-256. doi:10.1007/s004360050105
- [53] Valero A., Martín-Sánchez J., Reyes-Muelas E., Adroher F.J. 2000. Larval anisakids parasitizing the blue whiting, *Micromesistius poutassou*, from Motril Bay in the Mediterranean region of southern Spain. *Journal of Helminthology* 74: 361-364. doi:10.1017/S0022149X00000536
- [54] Strømnes E., Andersen K. 1998. Distribution of whaleworm (Anisakis simplex, Nematoda, Ascarido-

idea) L₃ larvae in three species of marine fish; saithe (*Pollachius virens* (L.)), cod (*Gadus morhua* L.) and redfish (*Sebastes marinus* (L.)) from Norwegian waters. *Parasitology Research* 84: 281-285. doi:10.1007/s004360050396

- [55] Kleinertz S., Klimpel S., Palm H.W. 2012. Parasite communities and feeding ecology of the European sprat (*Sprattus sprattus* L.) over its range of distribution. *Parasitology Research* 110:1147-1157. doi:10.1007/s00436-011-2605-z
- [56] Rokicki J. 1972. Larwy Anisakis sp. u śledzi Clupea harengus L. w Bałtyku [Anisakis sp. larvae in Clupea harengus L. living in the Baltic Sea]. Wiadomości Parazytologiczne 18: 89-98 (in Polish).
- [57] Grabda J. 1974. The dynamics of the nematode larvae, *Anisakis simplex* (Rud.) invasion in the southwestern Baltic herring (*Clupea harengus* L.). *Acta Ichthyologica et Piscatoria* 4: 3-21.
- [58] Rolbiecki L., Rokicki J. 2002. III-stage Anisakis simplex (Rudolphi, 1809) (Nematoda: Anisakidae) larvae in herring caught in autumn from the Polish part of the Vistula Lagoon. Acta Scientarum Polonorum, Piscaria 1: 105-110.
- [59] Szostakowska B., Myjak P., Wyszyński M., Pietkiewicz H., Rokicki J. 2005. Prevalence of anisakid nematodes in fish from Southern Baltic Sea. *Polish Journal of Microbiology* 54 (Suppl.): 41-45.
- [60] Sobecka E., Łuczak E., Więcaszek B., Antoszek A. 2011. Parasite community structure of cod from Bear Island (Barents Sea) and Pomeranian Bay (Baltic Sea). *Polish Polar Research* 32: 253-262. doi:10.2478/v10183-011-0016-6
- [61] Grabda J. 1976. Studies on the life cycle and morphogenesis of *Anisakis simplex* (Rudolphi, 1809) (Nematoda; Anisakdae) cultured in vitro. *Acta Ichthyologica et Piscatoria* 6: 119-141.
- [62] Myjak P., Szostakowska B., Wojciechowski J., Pietkiewicz H., Rokicki J. 1994. Anisakid larvae in cod from the southern Baltic Sea. Archive of Fishery and Marine Research 42:149-161.
- [63] Mehrdana F., Bahlool Q.Z.M., Skov J., Marana M.H., Sindberg D., Mundeling M., Overgaard B.C., Korbut R., Strøm S.B., Kania P.W., Buchmann K. 2014. Occurrence of zoonotic nematodes *Pseudoterranova decipiens, Contracaecum osculatum* and *Anisakis simplex* in cod (*Gadus morhua*) from the Baltic Sea. *Veterinary Parasitology* 205: 581-587. doi:10.1016/j.vetpar.2014.08.027
- [64] Nadolna K., Podolska M. 2014. Anisakid larvae in the liver of cod (*Gadus morhua*) L. from the Southern Baltic Sea. *Journal of Helminthology* 88: 237-246. doi:10.1017/S0022149X13000096
- [65] Rolbiecki L., Rokicki J. 2000. The occurrence of the nematodes Anisakis simplex pathogenic to man in pike-perch from the Vistula Lagoon, Poland. Wiadomości Parazytologiczne 46: 397-402.
- [66] Grabda J. 1981. Parasitic fauna of garfish Belone

belone (L.) from the Pomeramian Bay (Southern Baltic) and Its Origin. *Acta Ichthyologica et Piscatoria* 11: 75-85.

- [67] Zander C. D., Döring W. 1989. The role of gobies (Gobiidae, Teleostei) in the food web of the shallow habitats of the Baltic Sea. In: *Proceedings of the 21th European Marine Biology Symposium* (Eds. R.Z. Klekowski, E. Styczyńska-Jurewicz, L. Falkowski). Polish Academy of Sciences, Institute of Oceanology, Gdańsk: 499-508.
- [68] Zander C.D., Strohbach U., Groenewold S. 1993. The importance of gobies (Gobiidae, Teleostei) as hosts and transmitters of parasites in the SW Baltic. *Helgoländer Meeresuntersuchungen* 47: 81-111. doi:10.1007/BF02366186
- [69] Podolska M., Morozińska J. 1994. Anisakis simplex (larva III stage) in stickleback Gasterosteus aculeatus from the southern Baltic region. Wiadomości Parazytologiczne 40: 305-309 (in Polish with summary in English).
- [70] Køie M. 2001. Experimental infections of copepods and sticklebacks *Gasterosteus aculeatus* with small ensheathed and large third-stage larvae of *Anisakis simplex* (Nematoda, Ascaridoidea, Anisakidae). *Parasitology Research* 87: 32-36. doi:10.1007/s004360000288
- [71] Rolbiecki L., Janc A., Rokicki J. 2001. Stickleback as a potential paratenic host in the *Anisakis simplex* life cycle in the Baltic Sea: results of experimental infection. *Wiadomości Parazytologiczne* 47: 257-262.
- [72] Podolska M., Horbowy J., Wyszyński M. 2006. Discrimination of Baltic herring populations with respect to Anisakis simplex larvae infection. Journal of Fish Biology 68:1241-1256. doi:10.1111/j.0022-1112.2006.01004.x
- [73] Podolska M., Horbowy J. 2003. Infection of Baltic herring (*Clupea harengus membras*) with *Anisakis* simplex larvae, 1992–1999: a statistical analysis using generalized linear models. *ICES Journal of Marine Science* 60: 85-93. doi:10.1006/jmsc.2002.1323
- [74] Horbowy J., Podolska M. 2001. Modelling infection of Baltic herring (*Clupea harengus membras*) by larval Anisakis simplex. ICES Journal of Marine Science 58: 321–330. doi:10.1006/jmsc.2000.1013
- [75] Podolska M. 2009. Larwy Anisakis simplex w śledziach bałtyckich jako organizmy wskaźnikowe w badaniach biologicznych i rybackich. I. Śledź bałtycki i nicienie A. simplex: modele infekcji pasożytniczej i wskaźniki migracji ryb [Anisakis simplex larvae from the Baltic herring as tag organisms in biological and fisheries research. I. Baltic herring and A. simplex nematodes: the models of parasitic infection and fish migration tags]. Wiadomości Parazytologiczne 55: 201-210 (in Polish with summary in English).
- [76] Molina-Fernández D., Malagón D., Gómez-Mateos M., Benitez R., Martín Sánchez J. Adroher F.J. 2015.

Fishing area and fish size as risk factors of *Anisakis* infection (*Sardina pilchardus*) from Iberian waters, southwestern Europe. *International Journal of Food Microbiology* 203: 27-34.

doi:10.1016/j.ijfoodmicro.2015.02.024

- [77] Van Thiel P.H. 1962. Anisakiosis. *Parasitology* 52 (Suppl.): 16-17.
- [78] Pravettoni V., Primavesi L., Piantanida M. 2012. Anisakis simplex: current knowledge. European Annals of Allergy and Clinical Immunology 44: 150-156.
- [79] Bucci C., Gallotta S., Morra I., Fortunato A., Ciacci C., Iovino P. 2013. Anisakis, just think about it in an emergency! *International Journal of Infectious Diseases* 17: e1071- e1072. doi:10.1016/j.ijid.2013.05.008
- [80] Couture C., Measures L., Gagnon J., Desbiens C. 2003. Human intestinal anisakiosis due to consumption of raw salmon. *American Journal of Surgical Pathology* 27: 1167-1172.
- [81] Hochberg N.S., Hamer D.H., Hughes J.M., Wilson M.E. 2010. Anisakidosis: Perils of the Deep. *Clinical Infectious Diseases* 51: 806-812. doi:10.1086/656238
- [82] Buchmann K., Mehrdana F. 2016. Effects of anisakid nematodes Anisakis simplex (s.l.), Pseudoterranova decipiens (s.l.) and Contracaecum osculatum (s.l.) on fish and consumer health. Food and Waterborne Parasitology 4: 13-22. doi:10.1016/j.fawpar.2016.07.003
- [83] Mladineo I., Popović M., Drmić-Hofman I., Poljak V. 2016. A case report of Anisakis pegreffii (Nematoda, Anisakidae) identified from archival paraffin sections of a Croatian patient. BMC Infectious Disease 16: 42. doi:10.1186/s12879-016-1401-x
- [84] Nicuwenhuizen N., Lopata A.L., Jeebhay M.F., De'Broski R.H., Robins T.G., Brombacher F. 2006. Exposure to the fish parasite *Anisakis* causes allergic airway hyperreactivity and dermatitis. *Journal of Allergy and Clinical Immunology* 117: 1098-1105. doi:10.1016/j.jaci.2005.12.1357
- [85] Abattouy N., Valero A., Benajiba M.H., Lozano J., Martín-Sánchez J. 2011. Anisakis simplex s.l. parasitization in mackerel (Scomber japonicus) caught in the North of Morocco – prevalence and analysis of risk factors. International Journal of Food Microbiology 150:136-139.

doi:10.1016/j.ijfoodmicro.2011.07.026

- [86] Abattouy N., Valero A., Martín-Sánchez J., Peńalver M.C., Lozano J. 2012. Sensitization to Anisakis simplex species in the population of northern Morocco. Journal of Investigational Allergology and Clinical Immunology 22: 514-519.
- [87] Shamsi S., Butcher A.R. 2011. First report of human anisakidosis in Australia. *Medical Journal of Australia* 194: 199-200. doi:10.5694/j.1326-5377.2011.tb03772.x

- [88] Serrano-Moliner M., Morales-Suarez-Varela M., Adela Valero M.A. 2018. Epidemiology and management of foodborne nematodiasis in the European Union, systematic review 2000–2016, *Pathogens and Global Health* 112: 249-258. doi:10.1080/20477724.2018.1487663
- [89] Herrador Z., Daschner A., Perteguer M.J., Benito A. 2019. Epidemiological scenario of anisakidosis in Spain based on associated hospitalizations: the tipp of the iceberg. *Clinical Infection Diseases* 69: 69-76. doi:10.1093/cid/ciy853
- [90] Guardone L., Armani A., Nucera D., Costanzo F., Mattucci S., Bruschi F. 2018. Human anisakiasis in Italy: a retrospective epidemiological study over two decades. *Parasite* 25 41. doi:10.1051/parasite/2018034
- [91] Van Thiel P.H., Kuipers F.C., Roskam R.T. 1960. A nematode parasitic to herring, causing acute abdominal syndromes in man. *Tropical and Geographical Medicine* 2: 97-113.
- [92] Guz L., Studzińska M.B., Sadzikowski A.B., Gundłach J.L. 2005. Larwy Anisakis simplex w wędzonych śledziach [Anisakis simplex larval infestation in smoked herring]. Annales UMCS, sectio DD: 60, 88-94 (in Polish with summary in English).
- [93] Horbowy J., Podolska M., Nadolna-Ałtyn K. 2016. Increasing occurrence of anisakid nematodes in the liver cod (*Gadus morhua*) from the Baltic Sea: Does infection affect the condition and mortality of fish? *Fisheries Research* 179: 98-103. doi:10.1016/j.fishres.2016.02.011
- [94] Guz L., Studzińska M.B., Sadzikowski A.B., Gundłach J.L. 2005. Anisakioza [Anisakiosis]. Annales UMCS, sectio DD, 60: 74-87(in Polish with summary in English).
- [95] Bilska-Zając E., Lalle M., Różycki M., Chmurzyńska E., Kochanowski M., Karamon J., Sroka J., Pozio E., Cencek T. 2016. High prevalence of Anisakidae larvae in marketed frozen fillets of pink salmons (*Oncorhy nchus gorbuscha*). Food Control 68: 216-219. doi:10.1016/j.foodcont.2016.03.049
- [96] Aloia A., Carlomagno P., Gambardella M., Schiavo M., Pasquale V. 2011. Accidental endoscopic finding of *Anisakis simplex* in human colon. *Microbiologia Medica* 26: 209-211. doi:10.4081/mm.2011.2359
- [97] Baird F.J., Gasser R.B., Jabbar A., Lopata A.L. 2014. Foodborne anisakiosis and allergy. *Molecular* and Cellular Probes 28: 167-174. doi:10.1016/j.mcp.2014.02.003
- [98] Ivanovic J., Baltic M.Z., Boskovic M., Kilibarda N., Dokmanovic M., Markovic R., Janjic J., Baltic B. 2015. Anisakis infection and allergy in humans. Procedia Food Science 5: 101-104. doi:10.1016/j.profoo.2015.09.028
- [99] Madi L., Ali M., Legace-Wiens P., Duerksen D.R. 2013. Gastrointestinal manifestations and management of anisakiosis. *Canadian Journal of Gastroenterology*

27: 126-127. doi:10.1155/2013/427982

- [100] Carlin A.F., Abeles S., Chin N.A., Lin G.Y., Young M., Vinetz J.M. 2018. Case report: A common source outbreak of anisakidosis in the United States and postexposure prophylaxis of family collaterals. *The American Journal of Tropical Medicine and Hygiene* 99: 1219-1221. doi:10.4269/ajtmh.18-0586
- [101] Kasuya S., Hamano H., Izumi S. 1990. Mackerelinduced urticaria and *Anisakis. Lancet* 335: 665. doi:10.1016/0140-6736(90)90455-E
- [102] Daschner A., Cuellar C., Rodero M. 2012. The Anisakis allergy debate: does an evolutionary approach help? Trends in Parasitology 28: 9-15. doi:10.1016/j.pt.2011.10.001
- [103] Mattiucci S., Fazii P., De Rosa A., Paoletti M., Megna A.S., Glielmo A., De Angelis D., Costa A., Meucci C., Calvaruso V., Sorrentini I., Palma G., Bruschi F., Nascetti G. 2013. Anisakiosis and gastroallergic reactions associated with *Anisakis* pegreffii infection, Italy. *Emerging Infectious* Diseases 19: 496-499. doi:10.3201/eid1903.121017
- [104] Caballero M.L., Moneo I. 2004. Several allergens from *Anisakis simplex* are highly resistant to heat and pepsin treatments. *Parasitology Research* 93: 248-251. doi:10.1007/s00436-004-1099-3
- [105] Piccolo G., Manfredi M.T., Hoste L., Vercruysse J. 1999. Anisakidae larval infection in fish fillets sold in Belgium, *Veterinary Quarterly* 21: 66-67. doi:10.1080/01652176.1999.9694995
- [106] Rodríguez-Mahillo A.I., González-Muňoz M., Moneo I., Solas T.M., Mendizabal A., De Las Heras Ch., Tejada M. 2008. Allergenic properties and cuticle microstructure of *Anisakis simplex* L3 after freezing and pepsin digestion. *Journal of Food Protection* 71: 2578-2581. doi:10.4315/0362-028X-71.12.2578
- [107] Moneo I., Carballeda-Sangiao N., González-Muňoz M. 2017. New perspectives on the diagnosis of allergy to *Anisakis* spp. *Current Allergy and Asthma Reports* 17: 27.

doi:10.1007/s11882-017-0698-x

- [108] Gonzalez-Munoz M., Luque R., Nauwelaers F., Moneo I. 2005. Detection of Anisakis simplexinduced basophil activation by flow cytometry. Cytometry Part B (Clinical Cytometry) 68B: 31-36. doi:10.1002/cyto.b.20070
- [109] Armentia A., Martín-Gil F.J., Pascual C., Martín-Esteban M., Callejo A., Martínez C. 2006. Anisakis simplex allergy after eating chicken meat. Journal of Investigational Allergology and Clinical Immunology 16: 258-263.
- [110] Ańíbarro B., Seoane F.J. 1998. Occupational conjunctivitis caused by sensitization to Anisakis simplex. Journal of Allergy and Clinical Immunology 102: 331-332. doi:10.1016/S0091-6749(98)70108-3
- [111] Lopata A.L., Jeebhay M.F. 2013. Airborne seafood allergens as a cause of occupational allergy and

asthma. Current Allergy and Asthma Reports 13: 288-297. doi:10.1007/s11882-013-0347-y

- [112] Nieuwenhuizen N., Lopata A.L. 2014. Allergic reactions to Anisakis found in fish. Current Allergy and Asthma Reports 14: 455 doi:10.1007/s11882-014-0455-3
- [113] Ivanovic J., Baltic M.Z., Boskovic M., Kilibarda N., Dokmanovic M., Markovic R., Janjic J., Baltic B. 2017. Anisakis allergy in human. Trends in Food Science & Technology 59: 25-29. doi:10.1016/j.tifs.2016.11.006
- [114] Moneo I., Caballero M.L., González-Muňoz M., Rodríguez-Mahillo A.I., Rodríguez-Perez R., Silva A. 2005. Isolation of a heat-resistant allergen from the fish parasite *Anisakis simplex*. *Parasitology Research* 96: 285-289.

doi:10.1007/s00436-005-1362-2

- [115] Rodríguez-Mahillo A.I., González-Muñoz M., de las Heras C,. Tejada M., Moneo I. 2010. Quantification of *Anisakis simplex* allergens in fresh, longterm frozen, and cooked fish muscle. *Foodborne Pathogens and Disease* 7: 967-973. doi:10.1089/fpd.2009.0517
- [116] Vidaček S., De Las Heras C., Solas M.T., García M.L., Mendizábal A., Tejada M. 2011. Viability and antigenicity of *Anisakis simplex* after conventional and microwave heating at fixed temperatures. *Journal* of Food Protection 74: 2119-2126. doi:10.4315/0362-028X.JFP-11-108
- [117] Toro C., Caballero M.L., Baquero M., García-Samaniego J., Casado I., Rubio M., Moneo I. 2004. High prevalence of seropositivity to a major allergen of *Anisakis simplex*, Ani s 1, in dyspeptic patients. *Clinical and Diagnostic Laboratory Immunology* 11: 115-118. doi:10.1128/CDLI.11.1.115-118.2004
- [118] Anadón A.M., Romarís F., Escalante M., Rodríguez E., Gárate T., Cuéllar C., Ubeira F.M. 2009. The Anisakis simplex Ani s 7 major allergen as an indicator of true Anisakis infections. Clinical and Experimental Immunology 156: 471-478. doi:10.1111/j.1365-2249.2009.03919.x
- [119] Moneo I., Caballero M.L., Rodriguez-Perez R., Rodriguez-Mahillo A.I., Gonzalez-Muńoz M. 2007. Sensitization to the fish parasite Anisakis simplex: clinical and laboratory aspects. Parasitology Research 101: 1051-1055. doi:10.1007/s00436-007-0587-7
- [120] Lorenzo S., Iglesias R., Leiro J., Ubeira F.M., Ansotegui I., García M., Fernández de Corres L. 2000. Usefulness of currently available methods for the diagnosis of *Anisakis simplex* allergy. *Allergy* 55: 627-633. doi:10.1034/j.1398-9995.2000.00471.x
- [121] Carballeda-Sangiao N., Rodríguez-Mahillo A.I., Puente S., Gutiérrez M.T., Moneo I., González-Munoz M. 2014. Anisakis/Ascaris IgE ratio improves specificity for the diagnosis of Anisakis simplex sensitization intravellers and immigrants. Acta

Tropica 138: 1-4.

doi:10.1016/j.actatropica.2014.05.020

- [122] Celano G.V., Paparella A., Fransvea A., Balzaretti C., Celano G. 2013. Rapid method for detection of Anisakidae larvae in marine fishes, based on UV transillumination. *International Journal of Bio-science, Biochemistry and Bioinformatics* 3: 392-394. doi: 10.7763/IJBBB.2013.V3.240
- [123] Podolska M., Nadolna-Ałtyn K., Pawlak J., Różycki M., Pękala-Safińska A. 2019. Wykrywanie i identyfikacja nicieni Anisakidae – metody niedestrukcyjne [Detection and identification of Anisakidae nematodes – nondestructive methods]. In: Zasady dobrej praktyki w przetwórstwie rybnym (Eds. M. Różycki, M. Podolska). Państwowy Instytut Weterynaryjny – Państwowy Instytut Badawczy, Puławy: 100-108 (in Polish).
- [124] Różycki M., Bilska-Zając E., Kochanowski M., Grądziel-Krukowska K., Karamon J., Cencek T. 2019. Wykrywanie i identyfikacja nicieni Anisakidae – metody destrukcyjne [Detection and identification of Anisakidae nematodes – destructive methods]. In: Zasady dobrej praktyki w przetwórstwie rybnym (Eds. M. Różycki, M. Podolska). Państwowy Instytut Weterynaryjny – Państwowy Instytut Badawczy, Puławy: pp. 108-113 (in Polish).
- [125] Food and Drug Administration (FDA) 2011. Fish and Fishery Products Hazards and Controls Guidance 4th Edition Washington, DC: Department of Health and Human Services, Food and Drug Administration, Center for Food Safety and Applied Nutrition, downloads/Food/GuidanceRegulation/

UCM251970.pdf

- [126] Buchmann K., Kania P. 2012. Emerging *Pseudoterranova decipiens* (Krabbe, 1878) problems in Baltic cod, *Gadus morhua* L., associated with grey seal colonization of spawning grounds. *Journal of Fish Disease* 35: 861-866. doi:10.1111/j.1365-2761.2012.01409.x
- [127] Torres P., Jercic M.I., Weitz J.C., Dobrew E.K., Mercado R.A. 2007. Human Pseudoterranovosis, an emerging infection in Chile. *Journal of Parasitology* 93: 440-443. doi:10.1645/GE-946R.1
- [128] Ludovisi A., Di Felice G., Carballeda-Sangiao N., Barletta B., Butteroni C., Corinti S., Marucci G., González-Muňoz M., Pozio E., Gómez-Morales M.A. 2017. Allergenic activity of *Pseudoterranova decipiens* (Nematoda: Anisakidae) in BALB/c mice. *Parasites & Vectors* 10: 290. doi:10.1186/s13071-017-2231-4
- [129] Mehrdana F., Buchmann K. 2017. Excretory /secretory products of anisakid nematodes: biological and pathological roles. *Acta Veterinaria Scandinavica* 59: 42. doi:10.1186/s13028-017-0310-3
- [130] Podolska M., Pawlikowski B., Nadolna-Ałtyn K., Pawlak J., Komar-Szymczak K., Szostakowska B. 2019. How effective is freezing at killing *Anisakis* simplex, Pseudoterranova krabbei, and P. decipiens larvae? An experimental evaluation of timetemperature conditions. Parasitology Research 118: 2139-2147. doi:10.1007/s00436-019-06339-1

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