Short note

Blastocystis isolates from a dog and their owners presenting with chronic diarrhoea. Dogs as reservoirs of *Blastocystis*: research in Poland and worldwide.

Adam KACZMAREK¹, Anna ROCKA², Maria WESOŁOWSKA³, Elżbieta GOŁĄB⁴, Rusłan SAŁAMATIN^{1,5}

¹Faculty of Medicine, Collegium Medicum, Cardinal Stefan Wyszyński University in Warsaw, ul. Kazimierza Wóycickiego 1/3, 01-938 Warsaw, Poland

²W. Stefański Institute of Parasitology, Polish Academy of Sciences, ul. Twarda 51/55, 00-818 Warsaw, Poland ³Department of Biology and Medical Parasitology, Wrocław Medical University, ul. J. Mikulicza-Radeckiego 9, 50-345 Wrocław, Poland

⁴The National Institute of Public Health – National Institute of Hygiene, ul. Chocimska 24, 00-791 Warsaw, Poland

⁵Department of General Biology and Parasitology, Medical University of Warsaw, ul. Chałubińskiego 5, 02-004 Warsaw, Poland

Corresponding Author: Rusłan SAŁAMATIN; e-mail:rsalamatin@gmail.com

ABSTRACT. *Blastocystis* cf. *hominis* is an unicellular protozoan parasite commonly found in the gastrointestinal tract of humans and animals. *Blastocystis* is characterized by high morphological and genetic diversity. Studies based on the analysis of *Blastocystis* spp. small subunit ribosomal RNA genes (SSU rDNA) have identified 26 subtypes (ST) so far, including at least 10 isolated from humans (STs 1–9 and ST12). In 2017, stool samples from a dog and its two owners living in Gdynia, Poland were examined; all three were suffering from chronic diarrhoea. In addition, 30 faecal samples were also examined from 30 dogs kept in one of Warsaw's hotels for animals. Stool specimens were analyzed using anaerobic cultivation at 37°C with a modified Jones' medium and molecular methods (PCR). Phylogenetic analysis using Bayesian inference was performed. Vacuolar forms of *Blastocystis* were identified in the stool samples of the dog and its owners; *Blastocystis* were not detected in any sample from the dogs living in the animal hotel. Based on the phylogenetic analysis, the obtained isolates were classified as subtype ST3 (for Owner 1) and subtype ST7 (for Owner 2 and the dog). To the best of our knowledge, the present study is the first to evaluate the presence of *Blastocystis* in canines in Poland, including domestic dogs.

Keywords: Blastocystis cf. hominis, dog, zoonosis, PCR

Introduction

Blastocystis cf. *hominis* is an unicellular protozoan parasite commonly found in the gastrointestinal tract of humans and animals [1,2]. Its worldwide distribution means it is one of the most frequentlydetected protozoans in humans [3]; however, its occurrence depends on the geographic region and various socioeconomic and environmental factors. Epidemiological data demonstrates that prevalence of *Blastocystis* infection varies from 20% in developed countries [4] up to 100% in developing countries [5]. *Blastocystis* is characterized by high morphological and genetic diversity. Studies based on the analysis of *Blastocystis* spp. small subunit ribosomal RNA genes (SSU rDNA) have identified 26 subtypes (ST) so far, including at least 10 isolated from humans (STs 1–9 and ST12). Most of the STs isolated from humans are ST1 through ST4; subtypes ST5 through ST9, and ST12, considered animal, are less common in humans [4,6,7].

Zoonotic transmission of *Blastocystis* (from animals to humans) can occur through several routes: direct transmission (poor hand hygiene) or *via* consumption of contaminated water or food. Research carried out in zoos in the United Kingdom, Philippines and Australia have confirmed the presence of the same genotypes of *Blastocystis* in animals and their carers [7–10]. In addition, Lee et al. [11] identified the same *Blastocystis* subtype in humans and their animals which draw water from a river.

The impact of *Blastocystis* on human health is still unclear, but many reports suggest it acts as a diarrheagenic agent [12].

In this paper we describe the first case of *Blastocystis* cf. *hominis* ST7 in a dog in Poland and the results of a study of 30 dogs kept in a hotel for animals.

Materials and Methods

In 2017, stool samples from a dog and its two owners living in Gdynia were examined; all three were suffering from chronic diarrhoea. In addition, 30 faecal samples were also examined from 30 dogs kept in one of Warsaw's hotels for animals.

Identification was performed using the microscopic method and by anaerobic cultivation at 37°C using modified Jones' medium supplemented with 10% horse serum [13]. The cultures were examined for the vacuolar form after 48 hours using light microscopy [14].

Any positive cultures were sub-cultured onto new medium and incubated for 48 hours, after which the cell pellet was separated by centrifugation at 70×g for one minute. DNA isolation from the cell pellet was performed using the Genomic Mini kit (A&A Biotechnology, Gdynia, Poland). A small subunit ribosomal RNA gene (SSU rDNA) fragment of about 560 bp was amplified by PCR method, with forward primer RD5 (5'-ATCTGGTTGATC CTGCCAGT-3') [15] and reverse primer BhRDr (5'-GAGCTTTTTAACTGCAACAACG-3') [16].

All PCR products were sequenced and the obtained sequences were compared to *Blastocystis* sequences deposited in GenBank. Phylogenetic analysis using Bayesian inference was performed using MrBayes 3.2.7a [17,18] including 24 reference sequences representing *Blastocystis* cf. *hominis* ST1–ST9 subtypes [8,14,19]. *Proteromonas lacertae* (GenBank: U37108) was used as an outgroup. *Blastocystis* subtype nomenclature follows that of Stensvold et al. [20]. The sequences reported in this paper were deposited in the GenBank database with the accession numbers

Figure 1. Vacuolar form of *Blastocystis* cf. *hominis* from culture (Owner 1 sample). Wet mount, Lugol's iodine. Scale bar 100 µm.

MW346667-MW346669.

Results and Discussion

Vacuolar forms of *Blastocystis* were identified in the stool samples of the dog and its owners from Gdynia, Poland (Fig. 1). However, *Blastocystis* were not detected in any sample from the dogs living in the animal hotel. Based on the phylogenetic analysis, the obtained isolates were classified as subtype ST3 (for Owner 1) and subtype ST7 (for Owner 2 and the dog) (Figs. 2–3).

Both subtypes have previously been reported in Poland. *Blastocystis* cf. *hominis* ST3 is considered to be typically found in humans. In Poland, subtype

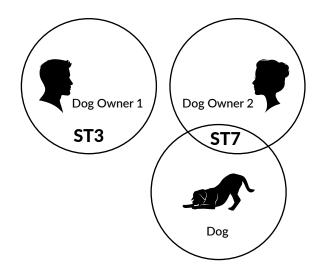


Figure 2. *Blastocystis* subtypes confirmed in the tested samples in this study

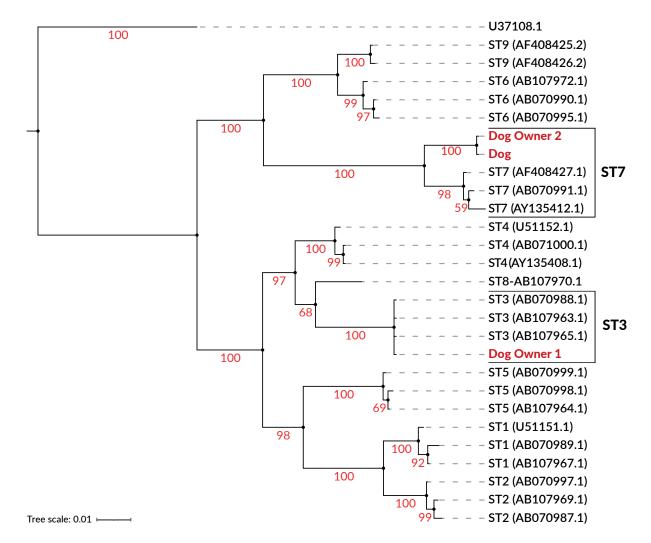


Figure 3. Bayesian inference tree based on fragment of sequences (536 bp and 548 bp) obtained at the small-subunit rRNA gene (SSU rDNA) of *Blastocystis* isolates of the present study, performed using MrBayes 3.2.7a [17,18]. The Bayesian posterior probabilities are shown adjacent to branch nodes.

ST3 was first detected in humans by Kotłowski [21] and later by Kaczmarek et al. [14,22]; Sałamatin et al. [23,24]; Wesołowska et al. [25,26]; its presence has also been noted by other researchers [27–32].

Blastocystis cf. *hominis* ST7 is considered typical for birds. While it was detected for the first time in Poland in chickens [33,34], it was later identified in humans [14,22,25,29–31].

The first animal detection of *Blastocystis* in Poland was made in 2014 in chickens [33–35]. Since then, *Blastocystis* has been detected in mammals at the Wrocław Zoo [36] and in pigs [32].

To the best of our knowledge, the present study is the first to evaluate the presence of *Blastocystis* in canines in Poland, including domestic dogs.

However, the presence of *Blastocystis* cf. *hominis* ST3 has been reported in dogs in Denmark [9] and Italy [37], and subtypes ST2 and ST10 in

France [38]. *Blastocystis* has also been observed in dogs in several countries in Asia, South America USA and in Australia.

In contrast, no *Blastocystis* infection has been reported in dogs in Spain or Greece or in Japan [39–42] and no reports on *Blastocystis* infection in dogs from Africa have been published (Fig. 4, Table 1).

The results of our present study are consistent with those of Wang et al. [43] and Ruaux et al. [44], who propose that dogs cannot act as natural hosts for *Blastocystis* due to there being such a low percentage of infection.

In conclusion, animals, that live in close contact with humans, such as dogs, should be considered as potential reservoirs of *Blastocystis*, however, due to their low prevalence of *Blastocystis*, dogs do appear to not pose such an important threat. Nevertheless,

Location	Subtypes (STs) identified	References
Argentina	Unknown	[45]
Australia	Unknown	[46,47]
Australia	ST1, ST3, ST4	[43,48]
Brazil	Unknown	[49–52]
Cambodia	ST2	[43]
Chile	Unknown	[53,54]
Colombia	ST2	[6,52]
Colombia	Unknown	[55]
Denmark	ST3	[2]
France	ST2, ST10	[38]
India	ST1, ST4, ST5, ST6	[43]
Iran	ST2, ST3, ST4, ST7, ST8, ST10	[58]
Iran	Unknown	[59–61]
Italy	ST3	[37]
Pakistan	Unknown	[62]
Philippines	Unknown	[56]
Philippines	ST1, ST2, ST3, ST4, ST5	[57]
Poland	ST7	This study
Thailand	ST5	[47]

Table 1. Summary of published studies/reports onBlastocystis spp. in dog

larger studies are needed to more accurately evaluate the problem presented by the zoonotic reservoir of *Blastocystis* in animals living close to humans.

References

- [1] Stenzel D.J., Boreham P.F.L. 1996. *Blastocystis hominis* revisited. *Clinical Microbiology Reviews* 9: 563-584.
- [2] Stensvold C.R., Alfellani M.A., Nørskov-Lauritsen S., Prip K., Victory E.L., Maddox C. et al. 2009. Subtype distribution of *Blastocystis* isolates from synanthropic and zoo animals and identification of a new subtype. *International Journal for Parasitology* 39: 473-479. doi:10.1016/j.ijpara.2008.07.006
- [3] Tan K.S.W. 2008. New insights on classification, identification, and clinical relevance of *Blastocystis* spp. *Clinical Microbiology Reviews* 21: 639-665. doi:10.1128/cmr.00022-08
- [4] Cian A., El Safadi D., Osman M., Moriniere R., Gantois N., Benamrouz-Vanneste S. et al. 2017. Molecular epidemiology of *Blastocystis* sp. in various animal groups from two French zoos and evaluation of potential zoonotic risk. *PLoS ONE* 12: e0169659. doi:10.1371/journal.pone.0169659
- [5] El Safadi D., Gaayeb L., Meloni D., Cian A., Poirier P., Wawrzyniak I. et al. 2014. Children of Senegal River Basin show the highest prevalence of *Blastocystis* sp. ever observed worldwide. *BMC Infectious Diseases* 14: 164.

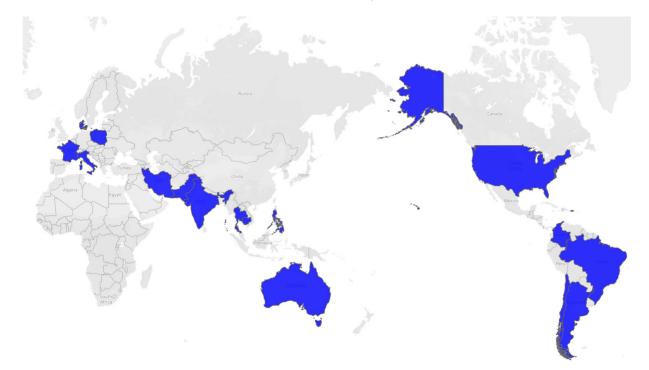


Figure 4. Geographical distribution of Blastocystis infection identified in dogs worldwide

doi:10.1186/1471-2334-14-164

- [6] Ramírez J.D., Sánchez L.V., Bautista D.C., Corredor A.F., Flórez A.C., Stensvold C.R. 2014. *Blastocystis* subtypes detected in humans and animals from Colombia. *Infection, Genetics and Evolution: Journal* of Molecular Epidemiology and Evolutionary Genetics in Infectious Diseases 22: 223-228. doi:10.1016/j.meegid.2013.07.020
- [7] Salim H.R., Kumar G.S., Vellayan S., Mak J.W., Khairul Anuar A., Init I. et al. 1999. *Blastocystis* in animal handlers. *Parasitology Research* 85: 1032-1033. doi:10.1007/s004360050677
- [8] Rivera W.L. 2008. Phylogenetic analysis of *Blastocystis* isolates from animal and human hosts in the Philippines. *Veterinary Parasitology* 156: 178-182. doi:10.1016/j.vetpar.2008.06.001
- [9] Stensvold C.R., Lewis H.C., Hammerum A.M., Porsbo L.J., Nielsen S.S., Olsen K.E.P. et al. 2009. *Blastocystis*: unravelling potential risk factors and clinical significance of a common but neglected parasite. *Epidemiology and Infection* 137: 1655-1663. doi:10.1017/s0950268809002672
- [10] Parkar U., Traub R.J., Vitali S., Elliot A., Levecke B., Robertson I. et al. 2010. Molecular characterization of *Blastocystis* isolates from zoo animals and their animal-keepers. *Veterinary Parasitology* 169: 8-17. doi:10.1016/j.vetpar.2009.12.032
- [11] Lee L., Chye T., Karmacharya B., Govind S. 2012. Blastocystis sp.: waterborne zoonotic organism, a possibility? Parasites and Vectors 5: 130. doi:10.1186/1756-3305-5-130
- [12] Skotarczak B. 2018. Genetic diversity and pathogenicity of *Blastocystis*. Annals of Agricultural and Environmental Medicine 25: 411-416. doi:10.26444/aaem/81315
- [13] Jones W.R. 1946. The experimental infection of rats with *Entamoeba histolytica*; with a method for evaluating the anti-amoebic properties of new compounds. *Annals of Tropical Medicine and Parasitology* 40: 130-140.

doi:10.1080/00034983.1946.11685270

- [14] Kaczmarek A., Gołąb E., Żarnowska-Prymek H., Rawska A., Jańczak D., Lewicki A. et al. 2017. Genetic diversity of *Blastocystis hominis sensu lato* isolated from humans in Poland [Zróżnicowanie genetyczne *Blastocystis hominis sensu lato* wyizolowanych od ludzi w Polsce]. *Przegląd Epidemiologiczny* 71: 539-546.
- [15] Clark C.G. 1997. Extensive genetic diversity in Blastocystis hominis. Molecular and Biochemical Parasitology 87: 79-83.

doi:10.1016/s0166-6851(97)00046-7

- [16] Scicluna S.M., Tawari B., Clark C.G. 2006. DNA barcoding of *Blastocystis*. *Protist* 157: 77-85. doi:10.1016/j.protis.2005.12.001
- [17] Huelsenbeck J.P., Ronquist F. 2005. Bayesian analysis of molecular evolution using MrBayes.

Statistical Methods in Molecular Evolution:183-226. doi:10.1007/0-387-27733-1 7

- [18] Miller M.A., Pfeiffer W., Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. 2010 Gateway Computing Environments Workshop, GCE 2010. doi:10.1109/gce.2010.5676129
- [19] Mattiucci S., Crisafi B., Gabrielli S., Paoletti M., Cancrini G. 2016. Molecular epidemiology and genetic diversity of *Blastocystis* infection in humans in Italy. *Epidemiology and Infection* 144: 635-646. doi:10.1017/s0950268815001697
- [20] Stensvold C.R., Suresh G.K., Tan K.S.W., Thompson R.C.A., Traub R.J., Viscogliosi E. et al. 2007. Terminology for *Blastocystis* subtypes – a consensus. *Trends in Parasitology* 23: 93-96. doi:10.1016/j.pt.2007.01.004
- [21] Kotłowski A. 2012. Blastocystoza próba oceny objawów klinicznych i skuteczności leczenia metronidazolem w przypadkach różnych stopni intensywności inwazji pasożytniczej oraz izolowanych genotypów *Blastocystis* sp. u Polaków powracających z tropiku i nieopuszczających kraju. *Annales Academie Medicae Gedanensis* 42 (Suppl. 3): 1-85 (in Polish).
- [22] Kaczmarek A., Wesołowska M., Gołąb E., Sałamatin R. 2019. *Blastocystis* spp. infection in young people in Poland. *Annals of Parasitology* 65 (Suppl.1): s59.
- [23] Salamatin R., Kaczmarek A., Rozej-Bielicka W., Wesolowska W., Mlocicki D., Golab E. 2016. Genotypes of *Blastocystis* isolated from Polish patients: a case of *Blastocystis hominis sensu lato* (subtype 6) infection. In Abstracts: *12th European Multicolloquium of Parasitology EMOP XII Turku*, *Finland, July 20–24th 2016*: P17.15.
- [24] Sałamatin R., Kaczmarek A., Rożej-Bielicka W., Cielecka D., Jańczak D., Lewicki A. et al. 2016. Genotype characterisation of *Blastocystis* isolates from Polish patients – preliminary results. *Annals of Parasitology* 62 (Suppl.): 93.
- [25] Wesołowska M., Frączkowski M., Rymer W., Janicki P., Poniewierka E., Puszyński G. 2019. Molecular subtyping of *Blastocystis* isolates from symptomatic and asymptomatic patients in Lower Silesia, Poland. *Annals of Parasitology* 65 (Suppl. 1): s72-73.
- [26] Wesolowska W., Kicia M., Szetela B., Kopacz Z., Salamatin R., Rymer W. et al. 2016. Prevalence of *Blastocystis hominis* among HIV-positive and HIVnegative patients in Poland. In Abstracts: 12th European Multicolloquium of Parasitology EMOP XII Turku, Finland, July 20–24th 2016: P6.11.
- [27] Bednarska M., Bajer A., Welc-Falçciak R., Pawełas A. 2015. Cyclospora cayetanensis infection in transplant traveller: a case report of outbreak. *Parasites and Vectors* 8: 411.

doi:10.1186/s13071-015-1026-8

- [28] Bednarska M., Jankowska I., Pawelas A., Piwczyńska K., Bajer A., Wolska-Kuśnierz B. et al. 2018. Prevalence of *Cryptosporidium*, *Blastocystis*, and other opportunistic infections in patients with primary and acquired immunodeficiency. *Parasitology Research* 117: 2869-2879. doi:10.1007/s00436-018-5976-6
- [29] Lepczyńska M., Dzika E., Stensvold C.R. 2016. Genetic diversity of *Blastocystis* spp. in the human population of the Olsztyn area. *Annals of Parasitology* 62 (Suppl.): s28.
- [30] Rudzińska M., Kowalewska B., Wąż P. 2019. Blastocystis subtypes distribution in patients performing routine coproscopy tests at the Department of Tropical Medicine and Epidemiology (Medical University of Gdańsk). Annals of Parasitology 65 (Suppl. 1): s69-70.
- [31] Rudzińska M., Kowalewska B., Wąż P., Sikorska K., Szostakowska B. 2019. *Blastocystis* subtypes isolated from travelers and non-travelers from the north of Poland – a single center study. *Infection, Genetics and Evolution* 75: 103926.
 - doi:10.1016/j.meegid.2019.103926
- [32] Rudzińska M., Kowalewska B., Szostakowska B., Grzybek M., Sikorska K., Świątalska A. 2020. First report on the occurrence and subtypes of *Blastocystis* in pigs in Poland using sequence-tagged-site pcr and barcode region sequencing. *Pathogens* 9: 1-14. doi:10.3390/pathogens9070595
- [33] Lewicki A., Rożej-Bielicka W., Sałamatin R. 2016. Blastocystis hominis s. l. ST6 – parasite of chickens – new zoonotic agent in Poland. Annals of Parasitology 62 (Suppl.): 203.
- [34] Kaczmarek A., Lewicki A., Dziedzic K., Sulecki K., Rożej-Bielicka W., Wesołowska M. et al. 2019. A survey of *Blastocystis* in domestic chickens from Poland and Madagascar. *Annals of Parasitology* 65 (Suppl. 1): s30.
- [35] Bobusia K., Gaweł A. 2014. Epizootiologia zarażeń pierwotniaczych Histomonas meleagridis, Tetratrichomonas gallinarum i Blastocystis spp. w stadach kur reprodukcyjnych mięsnych oraz indyków rzeźnych In: Aktualne problemy w patologii drobiu ze szczególnym uwzględnieniem chorób układu oddechowego (Ed. A. Wieliczko). Wydawnictwo Uniwersytetu Przyrodniczego, Wrocław: 80-82 (in Polish).
- [36] Wesołowska M., Paszta W., Michrowska A., Piekarska J., Wesołowska M., Gorczykowski M. et al. 2019. Molecular characterization of *Blastocystis* subtypes isolated from various mammalian groups living in Wrocław ZOO, Poland. *Annals of Parasitology* 65 (Suppl. 1): s47.
- [37] Gazzonis A.L., Marangi M., Zanzani S.A., Villa L., Giangaspero A., Manfredi M.T. 2019. Molecular epidemiology of *Blastocystis* sp. in dogs housed in Italian rescue shelters. *Parasitology Research* 118: 3011-3017. doi:10.1007/s00436-019-06424-5

- [38] Osman M., Bories J., El Safadi D., Poirel M.T., Gantois N., Benamrouz-Vanneste S. et al. 2015. Prevalence and genetic diversity of the intestinal parasites *Blastocystis* sp. and *Cryptosporidium* spp. in household dogs in France and evaluation of zoonotic transmission risk. *Veterinary Parasitology* 214: 167-170. doi:10.1016/j.vetpar.2015.09.015
- [39] Abe N., Nagoshi M., Takami K., Sawano Y., Yoshikawa H. 2002. A survey of *Blastocystis* sp. in livestock, pets, and zoo animals in Japan. *Veterinary Parasitology* 106: 203-212. doi:10.1016/p0204.4017(02)00050 x

doi:10.1016/s0304-4017(02)00050-x

- [40] Spanakos G., Papadogiannakis E., Kontos V., Menounos P., Velonakis E., Koutis C. et al. 2017. Molecular screening for *Blastocystis* sp. in canine faecal samples in Greece. *Journal of the Hellenic Veterinary Medical Society* 62: 216. doi:10.12681/jhvms.14852
- [41] Paulos S., Köster P.C., de Lucio A., Hernández-de-Mingo M., Cardona G.A., Fernández-Crespo J.C. et al. 2018. Pet dogs and cats do not seem to play a role as natural reservoirs of human blastocystosis in the province of Álava, Northern Spain. In Proceedings: 2nd International Blastocystis Conference. Bogotá, Colombia, October 11-12th 2018: PS13
- [42] Paulos S., Köster P.C., de Lucio A., Hernández-de-Mingo M., Cardona G.A., Fernández-Crespo J.C. et al. 2018. Occurrence and subtype distribution of *Blastocystis* sp. in humans, dogs and cats sharing household in northern Spain and assessment of zoonotic transmission risk. *Zoonoses and Public Health* 65: 993-1002. doi:10.1111/zph.12522
- [43] Wang W., Cuttell L., Bielefeldt-Ohmann H., Inpankaew T., Owen H., Traub R.J. 2013. Diversity of *Blastocystis* subtypes in dogs in different geographical settings. *Parasites and Vectors* 6: 215. doi:10.1186/1756-3305-6-215
- [44] Ruaux C.G., Stang B. V. 2014. Prevalence of *Blastocystis* in shelter-resident and client-owned companion animals in the US Pacific Northwest. *PLoS ONE* 9: e107496.

doi:10.1371/journal.pone.0107496

- [45] La Sala L.F., Leiboff A., Burgos J.M., Costamagna S.R. 2015. Spacial distribution of canine zoonotic endoparasites in Bahía Blanca, Argentina. *Revista Argentina de Microbiologia* 47: 17-24. doi:10.1016/j.ram.2014.12.006
- [46] Duda A., Stenzel D.J., Boreham P.F.L. 1998. Detection of *Blastocystis* sp. in domestic dogs and cats. *Veterinary Parasitology* 76: 9-17. doi:10.1016/s0304-4017(97)00224-0
- [47] Parkar U., Traub R.J., Kumar S., Mungthin M., Vitali S., Leelayoova S. et al. 2007. Direct characterization of *Blastocystis* from faeces by PCR and evidence of zoonotic potential. *Parasitology* 134: 359-367. doi:10.1017/s0031182006001582
- [48] Nagel R., Cuttell L., Stensvold C.R., Mills P.C.,

Bielefeldt-Ohmann H., Traub R.J. 2012. *Blastocystis* subtypes in symptomatic and asymptomatic family members and pets and response to therapy. *Internal Medicine Journal* 42: 1187-1195.

doi:10.1111/j.1445-5994.2011.02626.x

- [49] David É.B., Guimarães S., De Oliveira A.P., De Oliveira-Sequeira T.C.G., Bittencourt G.N., Nardi A.R.M. et al. 2015. Molecular characterization of intestinal protozoa in two poor communities in the State of São Paulo, Brazil. *Parasites and Vectors* 8: 1-12. doi:10.1186/s13071-015-0714-8
- [50] Moura R.G.F., de Oliveira-Silva M.B., Pedrosa A.L., Nascentes G.A.N., Cabrine-Santos M. 2018. Occurrence of *Blastocystis* spp. in domestic animals in triângulo mineiro area of Brazil. *Revista da Sociedade Brasileira de Medicina Tropical* 51: 240-243. doi:10.1590/0037-8682-0484-2016
- [51] Oliveira-Arbex A.P., David É.B., Tenório M. da S., Cicchi P.J.P., Patti M., Coradi S.T. et al. 2020. Diversity of *Blastocystis* subtypes in wild mammals from a zoo and two conservation units in southeastern Brazil. *Infection, Genetics and Evolution* 78: 104053. doi:10.1016/j.meegid.2019.104053
- [52] Jiménez P.A., Jaimes J.E., Ramírez J.D. 2019. A summary of *Blastocystis* subtypes in North and South America. *Parasites and Vectors* 12: 376. doi:10.1186/s13071-019-3641-2
- [53] Javier López D., Katia Abarca V., Patricio Paredes M., Elisa Inzunza T. 2006. Parasites intestinales en caninos y felinos con cuadros digestvos en Santiago, Chile. Consideracionesen Salud Pública [Intestinal parasites in dogs and cats with gastrointestinal symptoms in Santiago, Chile]. *Revista Medica de Chile* 134: 193-200 (in Spanish with summary in English)). doi:10.4067/s0034-98872006000200009
- [54] Opazo A., Barrientos C., Sanhueza A.M., Urrutia N., Fernández I. 2019. Fauna parasitaria en caninos (*Canis lupus familiaris*) de un sector rural de la región central de Chile. *Revista de Investigaciones Veterinarias del Perú* 30: 330-338 (in Spanish with summary in English).

doi:10.15381/rivep.v30i1.15683

[55] González A.C., Giraldo J.C. 2015. Prevalencia de

parásitos intestinales zoonóticos en caninos (*Canis lupus familiaris*) del área urbana del municipio de Coyaima (Tolima). *Revista Med* 23: 24 (in Spanish). doi:10.18359/rmed.1743

- [56] Tan M.A.V., Rivera W. 2008. Genotypic characterization of *Blastocystis* isolates in the Philippines. *Philippine Journal of Veterinary Medicine* 45: 86-94.
- [57] Belleza M.L.B., Reyes J.C.B., Tongol-Rivera P.N., Rivera W.L. 2016. Subtype analysis of *Blastocystis* sp. isolates from human and canine hosts in an urban community in the Philippines. *Parasitology International* 65: 291-294.

doi:10.1016/j.parint.2016.02.009

- [58] Mohammadpour I., Bozorg-Ghalati F., Gazzonis A.L., Manfredi M.T., Motazedian M.H., Mohammadpour N. 2020. First molecular subtyping and phylogeny of *Blastocystis* sp. isolated from domestic and synanthropic animals (dogs, cats and brown rats) in southern Iran. *Parasites and Vectors* 13: 1-11.doi:10.1186/s13071-020-04225-9
- [59] Daryani A., Barmaki N., Ettehad G., Sharif M., Dehghan M., Nemati A. et al. 2006. A cross-sectional study of *Blastocystis hominis* in primary schoolchildren, Northwest Iran. *International Journal of Tropical Medicine* 1: 53-57.
- [60] Ithoi I., Jali A., Mak J.W., Wan Sulaiman W.Y., Mahmud R. 2011. Occurrence of *Blastocystis* in water of two rivers from recreational areas in Malaysia. *Journal of Parasitology Research* 2011. doi:10.1155/2011/123916
- [61] Mohaghegh M.A., Vafaei M.R., Ghomashlooyan M., Azami M., Falahati M., Azadi Y. et al. 2018. A wide diversity of zoonotic intestinal parasites in domestic and stray dogs in rural areas of Kermanshah province, Iran. *Tropical Biomedicine* 35: 82-90.
- [62] Hadi R., Khalil B., Khan N., Ibrahim F., Qureshi A. 2017. The prevalence of *Blastocystis* sp. in various animal hosts in captivity at Karachi Zoo. *FUUAST Journal of Biology* 7: 113-116.

Received 24 October 2020 Accepted 10 December 2020