## Short note

# Anisakid nematodes in dolphins (Cetacea: Delphinidae) from the Baltic Sea area

### Leszek ROLBIECKI<sup>1</sup>, Tytus KUCZKOWSKI<sup>1</sup>, Joanna N. IZDEBSKA<sup>1</sup>, Jerzy ROKICKI<sup>1</sup>, Joanna DZIDO<sup>1</sup>, Iwona PAWLICZKA<sup>2</sup>

<sup>1</sup>Department of Invertebrate Zoology and Parasitology, Faculty of Biology, University of Gdańsk, ul. Wita Stwosza 59, 80-308 Gdańsk, Poland <sup>2</sup>Prof. Krzysztof Skóra Hel Marine Station, Institute of Oceanography, Faculty of Oceanography and Geography

<sup>2</sup>Prof. Krzysztof Skóra Hel Marine Station, Institute of Oceanography, Faculty of Oceanography and Geography, University of Gdańsk, ul. Morska 2, 84-150 Hel, Poland

Corresponding Author: Leszek Rolbiecki; e-mail: leszek.rolbiecki@ug.edu.pl

**ABSTRACT.** Dolphins are rarely observed in the Baltic Sea, making only sporadic appearances as a result of their migrations. The study included six specimens: four white-beaked dolphins *Lagenorhynchus albirostris* and two striped dolphins *Stenella coeruleoalba*. Their gastrointestinal tracts were found to contain the nematodes *Anisakis simplex* (in both species) and *Contracaecum* sp. (only in the white-beaked dolphins). This is the first record of Anisakidae nematodes in dolphins found in the Baltic Sea.

Keywords: Anisakis simplex, Contracaecum sp., Lagenorhynchus albirostris, marine mammals, parasites, Stenella coeruleoalba

#### Introduction

The Delphinidae (Cetacea: Odontoceti) is a family of marine mammals comprising 37 species [1], some of which are widely distributed [2]. Although the members of this group are not native to the Baltic Sea area, representatives of 10 species have been observed there at least once [3]. One of such taxon is the white-beaked dolphin Lagenorhynchus albirostris (Gray, 1846), associated with the cold waters of the Northern Atlantic [2]. It is the second most common cetacean species in the Baltic Sea, following the native harbour porpoise Phocoena phocoena (Linnaeus, 1758), and it is frequently observed in its waters. The first documented case of its presence originates from Sweden in 1844, while the first report in Polish waters was published in 1862, near Kołobrzeg [3]. In contrast, the striped dolphin Stenella coeruleoalba (Meyen, 1833), a widely-distributed species, inhabiting temperate to tropical seas; is rarely observed in the Baltic Sea and is considered to be more thermophilic. The first verified stranding of this mammal originates from 1987 from the Swedish coast, with later stranding reported from Poland, in the area of the Vistula Spit (included in the present study) from 1998 and 1999, and in Danish waters in 2001 [3].

Stranded individuals give the possibility of conducting highly valuable parasitological tests. The Delphinidae are predatory mammals with a rather wide trophic spectrum, and they feed mainly on fish, cephalopods and crustaceans, swallowing their prey whole [2,4,5]. Moreover, like other migratory aquatic animals [6–8] they are also highly-mobile, and often exhibit very wide distribution areas [2,4,5]. Thus, they can be hosts to many different parasites, contributing to their dispersal and circulation in the environment.

As dolphins are facing increasing threats from both direct human activity and ongoing climate change [9], there is a need for regular studies aimed at monitoring their health status, as well as identifying the causes and outcomes of the extensive migrations into their parasite distribution boundaries.

Catalog no.	Collection date	Sampling locality (landing)	Sex	Age	Length (cm)	Weight (kg)	Parasite	Habitat	No. of parasites
Lagenor	Lagenorhynchus albirostris	Ş							
1D	13.06.1989	Baltic Sea (Kołobrzeg)	Ц	+0	214.0	146.0	146.0 Contracaecum sp.	intestine	1 L*
2D	13.06.1989	Baltic Sea (Kołobrzeg)	М	1	196.0	114.0	114.0 Anisakis simplex	intestine	18 L4
4D	21.08.1995	Gulf of Gdańsk (Hel)	Ц	7	229.0	165.5	165.5 Anisakis simplex	stomach	1 3, 1 3
5D	19.09.1998	Baltic Sea (fishing area P11/12)	Ц	ND	231.0	130.4	Ι	I	I
Stenella	Stenella coeruleoalba								
6D	05.12.1998	Gulf of Gdańsk (Przebrno)	Μ	ND	187.0	75.0	75.0 Anisakis simplex	stomach	26 ♀♀, 68 ♂♂, 24 L4
7D	04.04.1999	Gulf of Gdańsk (Skowronki)	Μ	ND	187.0	56.5	I	I	I
*: stag	e not determined,	*: stage not determined, F: female, L: larvae, M: male, ND: no data	data						

342

#### **Materials and Methods**

Retrospective parasitological analyses of dolphins were conducted based on the material collected in the Laboratory of Parasitology and General Zoology, University of Gdańsk, and from material obtained from the study of marine mammals by the Hel Marine Station, University of Gdańsk in 1989–1999. The study material included four white-beaked dolphins and two striped dolphins from the Polish zone of the Baltic Sea waters (Table 1). All were dead when obtained: one was stranded (specimen no. 7D), and others were bycaught – two in pelagic trawl nets (4D and 5D) and three in bottom set nets (1D, 2D and 6D).

The gastrointestinal tracts, i.e. stomachs and intestines, were subjected to parasitological analysis. After dissecting the stomachs and intestines, the collected helminths were fixed in a mixture of glacial acetic acid and 40% formaldehyde (19:1), and then preserved in 70% ethanol. The nematodes were then cleared using glycerin for microscope examination [10,11].

#### **Results and Discussion**

Nematodes were found in three of the whitebeaked dolphins and one striped dolphin. Of these, *Anisakis simplex* (Rudolphi, 1809) (Ascaridida: Anisakidae) (138 specimens in both dolphin species) was predominant over *Contracaecum* sp. (Ascaridida: Anisakidae) (one specimen in a whitebeaked dolphin) (Table 1).

This is the first identification of these nematodes in dolphins as vagrant species in the Baltic Sea. Furthermore, it is the first record of the presence of *Contracaecum* in *L. albirostris* throughout its entire distribution area.

Other genera and species of Anisakidae nematodes have been recorded in earlier studies of dolphins, including five species of *Anisakis (A. berlandi* Mattiucci, Cipriani, Webb, Paoletti, Marcer, Bellisario, Gibson et Nascetti, 2014; *A. pegreffii* Campana-Rouget et Biocca, 1955; *A. nascetti* Mattiucci, Paoletti et Webb, 2009; *A. simplex, A ty pica* (Diesing, 1860)), *Pseudoterranova decipiens* (Krabbe, 1878) and *Contracaecum* spp. [e.g. 4,5,12–22]. However, as reflected in our present findings, *Contracaecum* nematodes have been rarely found to be parasites for this host group: thus far, specimens of nematodes identified as *Contracaecum* sp., larvae and adult stages, have been only found in *S. coeruleoalba* from the Mediterranean Sea [17]. Among marine mammals, pinnipeds are viewed as typical hosts for these nematodes [16].

In contrast, A. simplex is an Arctic-Boreal species with a wide range of hosts. Its final hosts are mammals, mainly marine Cetacea (e.g. Balenopteridae, Delphinidae, Kogiidae, Monodontidae, Physeteridae, Ziphiidae), but some Pinnipedia are also infected [21,23]. Eggs are introduced to the water with the faeces of infected mammals. Second stage (L2) or third stage (L3) larvae then develop and, after hatching, the L2 or L3 larvae are ingested by crustaceans, particularly by those of the Euphasiacea. Subsequently, the crustaceans are ingested by numerous fish species, particularly planktivorous species, and cephalopods, which in turn constitute an infection source for larger predatory fish, in which the parasites accumulate. After consumption, these fish and cephalopods are populated by L3 larvae, invasive for mammals, which then molt to form L4 larvae and adults [21,24–26].

As these nematodes are of zoonotic importance, being known to cause anisakiosis in humans [21,27–31]. It is important to understand their full host range, and to monitor their source of origin and possible dispersal routes and circulation in ecosystems. Such studies should also include its potential hosts, as well as its final hosts, of which relatively little is known.

Furthermore, it is important to recognize threats to the populations of increasingly rare marine mammals, which are facing a growing threat of extinction [9]. Global phenomena such as climate change are of key importance, as these influence a range of factors, resulting in reduced water salinity, changes in temperature, habitat degradation, and changes in the trophic structure and availability of food (e.g. distribution or abundance of prev). These factors have an averse effect on the health of marine mammals, resulting in disturbed immunity and greater susceptibility to disease [9,32,33], which in turn favors the development of parasitoses. mechanism of anisakiasis Although the development has been best characterized for humans [e.g. 29-31,34,35], studies have indicated the possibility of lesions occurring in dolphins, such as gastric ulceration, fibrosis, jejunitis, and even chronic granulomatous and ulcerative dermatitis [4,13,36–39]. Regarding the dolphins included in the present study, the genus Anisakis is known to

have a pathogenic effect on *S. coeruleoalba* [4,13,38,40–42].

It should be emphasized that such comprehensive research of the Anisakidae, examining its effects on different hosts from a variety of locations and habitats, utilizing the advances of numerous disciplines, fits perfectly into the One Health concept for achieving the optimal health of people, animals and the environment.

#### References

- Committee on Taxonomy. 2020. List of marine mammal species and subspecies. Society for Marine Mammalogy. www.marinemammalscience.org, accessed on 24.03.2021.
- [2] Jefferson T.A., Webber M.A., Pitman R.L. 2015. Marine mammals of the world. 2nd ed. Academic Press. doi:10.1016/C2012-0-06919-0
- [3] Kinze C.C., Schulze G., Skóra K.E., Benke H. 2011. Zahnwale als Gastarten in der Ostsee. *Meer und Museum* 23: 53-82.
- [4] Evans P.G.H., Collet A. 2009. Genus Lagenorhynchus. In: Whales, porpoises and dolphins – order *Cetacea*. Document 31. 16th ASCOBANS Advisory Committee Meeting, Brugge, Belgium: 724-727.
- [5] Evans P.G.H., Smeenk C.S. 2009. Genus Stenella. In: Whales, porpoises and dolphins – order Cetacea. Document 31. 16th ASCOBANS Advisory Committee Meeting, Brugge, Belgium: 715-719.
- [6] Rolbiecki L., Izdebska J.N., Pawliczka I. 2019. Digenetic trematode Ogmogaster antarcticus (Notocotylidae) in a fin whale Balaenoptera physalus (Balaenopteridae) stranded in the Baltic Sea. Diseases of Aquatic Organisms 132: 143-149. doi:10.3354/dao03315
- [7] Rolbiecki L., Izdebska J.N., Dzido J. 2020. The helminthofauna of the garfish *Belone belone* (Linnaeus, 1760) from the southern Baltic Sea, including new data. *Annals of Parasitology* 66: 237-241. doi:10.17420/ap6602.260
- [8] Dzido J., Rolbiecki L., Izdebska J.N., Bednarek R. 2020. Checklist of the parasites of European eel *Anguilla anguilla* (Linnaeus, 1758) (Anguillidae) in Poland. *Biodiversity Data Journal* 8: e52346. doi:10.3897/BDJ.8.e52346
- [9] Terracciano G., Fichi G., Comentale A., Ricci E., Mancusi C., Perrucci S. 2020. Dolphins stranded along the Tuscan Coastline (Central Italy) of the "Pelagos Sanctuary": a parasitological investigation. *Pathogens* 9: 612. doi:10.3390/pathogens9080612
- [10] Moravec F. 1994. Parasitic nematodes of freshwater fishes of Europe. Academia, Praha.
- [11] Rolbiecki L. 2010. Diversity of metazoan parasite communities in selected fish species from water basins with different degrees of anthropogenic stress.

*Oceanological and Hydrobiological Studies* 39 (Suppl. 2): 3-150.

- [12] Raga J.A. 1994. Parasitismus bei Cetacea. In: Handbuch der Säugetiere Europas. Meeressäugers. Wale und delphine – *Cetacea*. Einführung, *Monodontidae*, *Phocoenidae*, *Delphinidae*. Bd. 6/1A (Eds. D. Robineau, R. Duguy, M. Klima). Aula-Verlag, Wiesbaden: 132-179.
- [13] Abollo E., López A., Gestal C., Benavente P., Pascual S. 1998. Macroparasites in cetaceans stranded on the northwestern Spanish Atlantic coast. *Diseases of Aquatic Organisms* 32: 227-231. doi:10.3354/dao032227
- [14] Gibson D.I., Harris E.A., Bray R.A., Jepson P.D., Kuiken T., Baker J.R., Simpson V.R. 1998. A survey of the helminth parasites of cetaceans stranded on the coast of England and Wales during the period 1990-1994. *Journal of Zoology* 244: 563-574. doi:10.1111/j.1469-7998.1998.tb00061.x
- [15] Archer F.I., Perrin W.F. 1999. Stenella coeruleoalba. Mammalian Species 603: 1-9. doi:10.2307/3504476
- [16] Mattiucci S., Nascetti G. 2008. Advances and trends in the molecular systematics of anisakid nematodes, with implications for their evolutionary ecology and host-parasite co-evolutionary processes. *Advances in Parasitology* 66: 47-148. doi:10.1016/S0065-308X(08)00202-9
- [17] Aytemiz I., Dede A., Danyer E., Tonay A.M. 2012. Morphological identification of parasites found in the stomach contents of bycaught striped dolphins (*Stenella coeruleoalba*) from Turkish Eastern Mediterranean Sea coast. *Journal of the Black Sea / Mediterranean Environment* 18: 238-245.
- [18] Blažeković K., Ivana Pleić I. L., Đuras M., Tomislav Gomerčić T., Mladineo I. 2015. Three Anisakis spp. isolated from toothed whales stranded along the eastern Adriatic Sea coast. International Journal for Parasitology 45: 17-31. doi:10.1016/j.ijpara.2014.07.012
- [19] Galatius A., Kinze C.C. 2016. Lagenorhynchus albirostris (Cetacea: Delphinidae). Mammalian Species 48: 35-47. doi:10.1093/mspecies/sew003
- [20] Kuhn T., Cunze S., Kochmann J., Klimpel S. 2016. Environmental variables and definitive host distribution: a habitat suitability modelling for endohelminth parasites in the marine realm. *Scientific Reports* 6: 30246. doi:10.1038/srep3024
- [21] Mattiucci S., Cipriani P., Levsen A., Paoletti M., Nascetti G. 2018. Molecular epidemiology of *Anisakis* and Anisakiasis: An ecological and evolutionary road map. *Advances in Parasitology* 99: 93-263. doi:10.1016/bs.apar.2017.12.001
- [22] Gomes T.L., Quiazon K.M., Kotake M., Fujise Y., Ohizumi H., Itoh N., Yoshinaga T. 2021. *Anisakis* spp. in toothed and baleen whales from Japanese waters with notes on their potential role as biological tags.

*Parasitology International* 80: 102228. doi:10.1016/j.parint.2020.102228

[23] Skrzypczak M., Rokicki J., Pawliczka I., Najda K., Dzido J. 2014. Anisakids of seals found on the southern coast of Baltic Sea. *Acta Parasitologica* 59: 165-172.

doi:10.2478/s11686-014-0226-2

- [24] Køie M., Berland B., Burt M.D.B. 1995. Development to third-stage larvae occurs in the eggs of *Anisakis simplex* and *Pseudoterranova decipiens* (Nematoda, Ascaridoidea, Anisakidae). *Canadian Journal of Fisheries and Aquatic Sciences* 52 (Suppl. 1): 134-139. doi.org/10.1139/f95-519
- [25] Klimpel S., Palm H.W., Rückert S., Piatkowski U. 2004. The life cycle of *Anisakis simplex* in the Norwegian Deep (northern North Sea). *Parasitology Research* 94: 1-9. doi:10.1007/s00436-004-1154-0
- [26] Klimpel S., Palm H.W. 2011. Anisakid nematode (Ascaridoidea) life cycles and distribution: Increasing zoonotic potential in the time of climate change? In: Progress in Parasitology (Ed. H. Mehlhorn). Parasitology Research Monographs vol. 2. Springer Verlag, Berlin, Heidelberg: 201-222. doi:10.1007/978-3-642-21396-0 11
- [27] Ishikura H., Namiki M. (eds) 1989. Gastric anisakiasis in Japan. Epidemiology, diagnosis, treatment. Springer Verlag, Tokyo.
- [28] Ishikura H., Kikuchi K. (eds) 1990. Intestinal anisakiasis in Japan. Infected fish, seroimmunological diagnosis, and prevention. Springer Verlag, Tokyo.
- [29] Lymbery A.J., Cheah F.Y. 2007. Anisakid nematodes and anisakiasis. In: *Food-borne parasitic zoonoses. Fish and plant-borne parasites* (Eds. K.D. Murrell, F. Bernard). Springer Science+Business Media, LLC, New York: 185-207.
- [30] Morozińska-Gogol J. 2019. Anisakis spp. as etiological agent of zoonotic disease and allergy in European region – an overview. Annals of Parasitology 65: 303-314. doi:10.17420/ap6504.214
- [31] Kołodziejczyk L., Szostakowska B., Sobecka E., Szczucki K., Stankiewicz K. 2020. First case of human anisakiasis in Poland. *Parasitology International* 76: 102073. doi:10.1016/j.parint.2020.102073
- [32] Learmonth J.A., Macleod C.D., Santos M.B., Pierce G.J., Crick H.Q.P., Robinson R.A. 2006. Potential effects of climate change on marine mammals. *Oceanography and Marine Biology: An Annual Review* 44: 431-464.
- [33] Evans P.G.H., Pierce G.J., Panigada S. 2010. Climate change and marine mammals. *Journal of the Marine Biological Association of the United Kingdom* 90: 1483-1487. doi:10.1017/S0025315410001815
- [34] Baird F.J., Gasser R.B., Jabbar A., Lopata A.L. 2014. Foodborne anisakiasis and allergy. *Molecular and*

*Cellular Probes* 28: 167-174. doi:10.1016/j.mcp.2014.02.003

- [35] Guardone L., Armani A., Nucera D., Costanzo F., Mattiucci S., Bruschi F. 2018. Human anisakiasis in Italy: a retrospective epidemiological study over two decades. *Parasite* 25: 41. doi:10.1051/parasite/2018034
- [36] van Beurden S.J., IJsseldijk L.L., Cremers H.J.W.M., Gröne A., Verheije M.H., Begeman L. 2015. Anisakis spp. induced granulomatous dermatitis in a harbour porpoise Phocoena phocoena and a bottlenose dolphin Tursiops truncates. Diseases of Aquatic Organisms 112: 257-263. doi:10.3354/dao02818
- [37] Hrabar J., Bočina I., Gudan Kurilj A., Đuras M., Mladineo I. 2017. Gastric lesions in dolphins stranded along the Eastern Adriatic coast. *Diseases of Aquatic Organisms* 125: 125-139. doi:10.3354/dao03137
- [38] Pons-Bordas C., Hazenberg A., Hernandez-Gonzalez A., Pool R.V., Covelo P., Sánchez-Hermosin P., López A., Saavedra C., Fraija-Fernández N., Fernández M., Aznar F.J. 2020. Recent increase of ulcerative lesions caused by *Anisakis* spp. in cetaceans from the north-east Atlantic. *Journal of Helminthology* 94: e127.

doi:10.1017/S0022149X20000115

- [39] Schick L., IJsseldijk L.L., Grilo M.L., Lakemeyer J., Lehnert K., Wohlsein P., Ewers C., Prenger-Berninghoff E., Baumgärtner W., Gröne A., Kik M.J.L., Siebert U. 2020. Pathological findings in white-beaked dolphins (*Lagenorhynchus albirostris*) and Atlantic white-sided dolphins (*Lagenorhynchus acutus*) from the south-eastern North Sea. *Frontiers in Veterinary Science* 7. doi:10.3389/fvets.2020.00262
- [40] Fernàndez M., Aznar J., Balbuena J.A., Raga J.A. 1991. Parasites collected in the striped dolphin die-off in the Spanish Mediterranean. Proceedings of the Fifth Conference of the European Cetacean Society, 21-23 February 1991, Sandefjord, Norway: 101-103.
- [41] Carvalho V.L., Bevilaqua C.M.L., Iñiguez A.M., Mathews-Cascon H., Ribeiro F.B., Pessoa L.M.B., de Meirelles A.C.O., Borges J.C.G., Marigo J., Soares L., de Lima Silva F.J. 2010. Metazoan parasites of cetaceans off the northeastern coast of Brazil. *Veterinary Parasitology* 173: 116-122. doi:10.1016/j.vetpar.2010.06.023
- [42] Oliveira J.B., Morales J.A., González-Barrientos R.C., Hernández-Gamboa J., Hernández-Mora G. 2011. Parasites of cetaceans stranded on the Pacific coast of Costa Rica. *Veterinary Parasitology* 182: 319-328. doi:10.1016/j.vetpar.2011.05.014

Received 25 March 2021 Accepted 06 May 2021